# The utility of exome sequencing in understanding the genetic basis of chronic kidney

#### disease

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## Abstract

Chronic kidney disease (CKD) is characterized by a progressive loss of renal function that can culminate to end stage renal disease (ESRD), requiring renal replacement therapy, which includes hemodialysis and kidney transplant as modalities. CKD has a prevalence of ~11-13% and is associated with significant morbidity and mortality. The prognosis and treatment of CKD cases varies by specific etiologies, which include congenital anomalies of the kidney and urinary tract (CAKUT), steroid-resistant nephrotic syndrome (SRNS), chronic glomerulopathies, cystic dominant tubulointerstitial kidney disease kidnev disease. autosomal (ADTKD). nephrolithiasis/nephrocalcinosis, and renal tubulopathies, among others. However, many patients are classified as CKD of unknown etiology. There have been significant and continuing investigations showing the genetic basis of CKD, specifically of monogenic causation (single-gene causing disease). However, current routine practices only use phenotype-driven clinical panel testing, which only evaluates a fraction of known monogenic CKD genes associated with a specific phenotype. Exome sequencing (ES) is thus a beneficial tool for CKD cases that are clinically misdiagnosed, or unknown, rendering them undiagnosed by genetic panel investigations. Although the diagnostic yield of ES has been shown to vary (10-50%), no such study has demonstrated its effectiveness in Canada. As such, this study has examined a cohort of 94 Canadian families who have been undiagnosed by current routine methods of genetic testing through, ES. We have shown that ES has an additional 17% diagnostic rate. Within this group, 58% of cases were categorized as CKDu, but all received a specific clinical diagnosis once solved. Additionally, a molecular diagnosis had a clinical impact for all patients who have been diagnosed, with diagnostic confirmation/resolution/correction, in addition to guidance on family planning, were among the most prevalent clinical benefits of receiving a genetic diagnosis. This study has demonstrated the added benefit of ES, in both the research and clinical settings.

Keywords: chronic kidney disease, monogenic, exome sequencing

#### Introduction

CKD is defined as a progressive loss of renal function due to abnormalities in either kidney structure or function for more than three months, in addition to either decreased estimated glomerular filtration rate (eGFR) or presence of markers of kidney damage, which includes albuminuria, urine sediment abnormalities, history of kidney transplants, or abnormalities either due to tubular disorders, ones detected via histology, or structural ones detected by imaging<sup>1</sup>. eGFR is commonly used to describe excretory kidney function, and when below 60 mL/min/ 1.73m, indicates a reduction in kidney function<sup>1</sup>. End stage renal disease (ESRD) is the culmination of this progressive disease, which establishes the requirement of renal replacement therapy, either by kidney transplant or dialysis<sup>2</sup>. Globally, CKD has been estimated to have a prevalence of 11-13%, and is associated with high morbidity and mortality<sup>3</sup>. In Canada, this number has been estimated to equate to 12.5%, which represents ~3 million Canadians <sup>4</sup>. CKD is further classified into specific etiologies, which includes congenital anomalies of the kidney and urinary tract (CAKUT), chronic glomerulopathies, cystic kidney disease, autosomal dominant tubulointerstitial kidney disease (ADTKD), nephrolithiasis/nephrocalcinosis, and renal tubulopathies, among others <sup>5</sup>. CKD of unknown etiology (CKDu) is the classification of patients which have CKD with an unknown cause, and this has been estimated to account for 10-36% of cases of this disease <sup>6</sup>. Given that the specific etiology of disease is essential in the determination of interventions as well as patient prognosis, developing methods to diagnose patients with such an etiology is beneficial.

Kidney disease that is known to have a genetic cause is referred to as genetic kidney disease (GKD)<sup>5</sup>. GKD can be further classified by causation, including polygenic - disease causation due to the combined effects of multiple genes - and monogenic - disease causation due

to a singular gene. Within monogenic causation, classification of mode of inheritance can be made. Autosomal dominant (AD) disease refers to the necessity for a singular parental allele to be mutated; autosomal recessive (AR) disease refers to the necessity for both parental alleles to be mutated; and X-linked (XL) disease refers to the necessity for a mutation on the X-chromosome to occur. Each mode of inheritance results in variation in the manner in which a phenotype is expected and is tightly correlated to genotypic changes. Currently, it is estimated that there is a ~30% prevalence of monogenic causation in CKD cases with onset before 25 years of age, with variation dependent on etiologies<sup>5</sup>. Although monogenic causation is more highly established among the pediatric population, given that there is a fraction of adult-onset cases with a positive family history, several studies have evaluated a monogenic causation in these cases. Previous studies by our lab have found that 36% of CKD patients with adult-onset and positive family history have a monogenic cause<sup>7</sup>. Given the contribution of monogenic causation to CKD, as well as the cruciality of understanding case-specific etiologies, a genetic diagnosis is an essential aspect to the care for CKD patients.

While sanger sequencing used to be the preferred method of genetic testing, the advent of next generation sequencing techniques has allowed for increased utility and benefit of genetic testing in clinical practice<sup>8</sup>. Through this method, simultaneous sequencing occurs, either in a particular of associated with group genes known to be а phenotype (targeted-phenotype-associated gene panels), of all known coding regions in the human genome (exome sequencing (ES)), or of the entire human genome (genome sequencing (GS))<sup>8</sup>. In routine clinical practices, targeted-phenotype associated gene panels are employed. However, given that many CKD patients do not receive a clinical diagnosis, or receive a misdiagnosis, and that new genes that are known to be causing CKD are routinely being discovered, ES provides benefits,

since ES analysis can evaluate all genes being interrogated - including those that are considered candidate genes. Using ES, all coding genes of the patient are sequenced, which allow for analysis of all genes known to be causing all forms of CKD at once, and they may be re-evaluated upon rapid updating of known gene lists<sup>8</sup>.

Recent studies have established a diagnostic yield of 10-50% using ES <sup>9,10</sup>, however no such study has been performed in Canada. Given its high diagnostic yield, in addition to the significant monogenic contribution already established in CKD and the benefits of establishing a genetic diagnosis, we hypothesize that the implementation of ES in routine clinical practice will be advantageous. Therefore, the aim of this study is to describe the prevalence and distribution of genetic disease within each phenotype by understanding the utility of ES as well as its clinical implications by performing this analysis using a curated list of known monogenic causes of CKD, on a cohort of 94 families that were undiagnosed by clinical gene panel investigations, and then by summarizing and evaluating several clinical parameters thereafter.

#### **Materials and Methods**

The general workflow of this study is summarized in Figure 1.

#### **Patients**

A total of n= 94 families, including n=102 patients were included in this study. This cohort comprised a group of families that were either unable to receive a genetic diagnosis by phenotype-targeted gene panels or did not receive a diagnosis by this method (Fig 1). Reasons for a lack of diagnosis include the broad specificity of symptoms relating to CKD phenotypes, leading to misdiagnosis, as well as only testing in a fraction of known genes by this method. These patients were referred to the genetic clinic at Victoria Hospital according to criteria that

increases suspicion that they have a genetic form of CKD. The cohort is described in Figure 2, and demonstrates the distribution of etiologies (Fig 2).

# Genetic clinic

Patients are referred to the renal genetic clinic at Victoria Hospital by their nephrologists, if there is a suspicion that they may have GKD. Although certain criteria for inclusion were established (Fig 1), they were also taken on a case-by-case basis to determine whether there is a risk of GKD. During the consultation, Dr. Connaughton reviews their medical and family history, and determines whether the patient is a candidate for phenotype-targeted gene panels, and if ES is a more suitable alternative, provides them with this option. Patients that are recruited follow consent procedures to participate in research ES. All participants provided informed written consent and the protocol was approved by the research ethics board at Western University.

#### **DNA** extraction

Genomic DNA was isolated from whole blood or saliva samples using the following protocols. Upon isolation, samples are stored in the SouthWestern Ontario Disease Genetic Biobank (SWORD-GEN Biobank). n=363 patients have been or are currently being investigated for GKD, either by ES or other methods.

#### Isolation of genomic DNA from whole blood

#### Cell Lysis

- Add 3mls of whole blood to a 15ml centrifuge tube containing 9ml of red blood cell (RBC) lysis solution. Invert to mix and incubate for 5 minutes at room temperature. Invert again at least once during incubation.
- 2. Centrifuge at 2000x g for 2 minutes to pellet the white blood cells (WBC).

- 3. Pour off the RBC lysis supernatant leaving behind the WBC pellet and drain tube for at least 10 seconds on clean absorbent paper. Less than 1  $\mu$ l residual liquid should remain.
- 4. Vortex the tube vigorously to resuspend the WBCs in the residual liquid.
- 5. Add 3ml of cell lysis solution to resuspend the cells.
- 6. To lyse the cells, vortex on high speed for 10 seconds. Usually no incubation is required; however, if cells clumps are visible, incubate at 37°C until the solution is homogenous. Samples are stable in cell lysis solution for at least 2 years at room temperature.

Protein precipitation

- 1. Cool samples to room temperature by placing samples on ice for 3 minutes.
- 2. Add 1 ml of protein precipitation solution to the cell lysate.
- 3. Vortex at high speed for 20 seconds to mix the protein precipitation solution uniformly with the cell lysate.
- 4. Centrifuge at 2000x g for 5 minutes. The precipitated protein should form a tight brown pellet. If the protein pellet is not tight, repeat step 3, followed by incubation on ice for 5 minutes and repeat step 4.

# DNA precipitation

- Pour the supernatant containing the DNA (leaving behind the protein pellet) into a clean 15ml centrifuge tube containing 3ml 100% isopropanol (2-propanol).
- Mix the sample by inverting gently 50 times until the white threads of DNA form visible clumps.
- 3. Centrifuge at 2000x g for 3 minutes. The DNA will be visible as a small white pellet.

- Pour off supernatant and drain tube briefly on clean absorbent paper. Add 3ml 70% ethanol and invert tube several times to wash the DNA pellet.
- Centrifuge at 2000x g for 1 minute. Carefully pour off the ethanol. The pellet may be loose so pour slowly.
- Invert and drain the tube on clean absorbent paper and allow to air dry for 10-15 minutes.

# DNA hydration

- Add 250µl of DNA hydration solution (250 µl will give a concentration of 400 µg/ml if the total yield is 100µg DNA). Note, DNA hydration solution consists of 10 mM tris, 1 mM EDTA with a pH of 7-8.
- Rehydrate DNA by incubating at 65°C for 1 hour or overnight at room temperature. If possible, tap the tube periodically to aid dispersing the DNA.
- 3. Samples may be centrifuged briefly and then transferred to storage tubes.
- 4. Store DNA at 4°C or -20 to -80°C for long term storage.

# Isolation of genomic DNA from saliva samples (using Orageneâ Discover DNA collection kit)

- 1. Incubate the spit sample at 50°C overnight.
- 2. Transfer the sample to a 15ml centrifuge tube.
- 3. Add 1ml of PrepIT-L2P solution to the tube and invert several times.
- 4. Place on ice for 10 minutes.
- 5. Centrifuge at room temperature for 5 minutes at 2000x g.
- 6. Carefully transfer the clear supernatant containing the DNA to a 15ml centrifuge tube containing 3mls of 100% ethanol. Invert 10 times to precipitate the DNA.

- Allow the sample to stand at room temperature for 10 minutes to allow the DNA to precipitate.
- 8. Centrifuge for 2 minutes at 2000x g.
- 9. Pour off supernatant allowing the DNA pellet to remain.
- 10. Add 1ml of 70% ethanol and gently invert to wash the DNA pellet. Allow to stand at room temperature for 1 minute.
- 11. Centrifuge for 2 minutes at 2000x g and pour off excess ethanol.
- **12**. Allow to stand at room temperature for 15 minutes to allow for complete evaporation of the ethanol.
- 13. Add 100µl of DNA hydration solution to dissolve the DNA pellet. Vortex for 5 seconds to dissolve.
- 14. Incubate at 37°C overnight on the shaker.
- 15. Store DNA at 4°C or -20 to -80°C for long term storage.

#### Exome sequencing, McGill 1-4

Purified genomic DNA was sent to the McGill Genome Centre at McGill University for whole exome sequencing using protocols for library preparation, exome capture, and next generation sequencing, according to the NxSeq AmpFREE low DNA Library Kit. 4 batches were used for this study, each group being numbered according to the batch that they were sent to McGill (1-4). FASTQ files were downloaded and processed using a custom automated workflow in CLC Genomics Workbench version 8.51 (CLC Bio, Aarhus, Denmark) for sequence alignment to our list of 735 genes known to be causing chronic kidney disease compiled via literature review in the lab (mapped to human genome build GRCh37/hg19), variant calling (generation of .VCF files), and target region coverage statistics (generation of .BAM files).

Variant annotation was conducted by using ANNOVAR with a customized Script. ANNOVAR is a functional annotation pipeline for genetic variants that allows for downloading sets of reference databases and *in silico* functional prediction, customize filtering strategies and calculate genetic risk score. Here, downloaded materials included: databases (RefSeq (hg19), dbSNP, gnomAD, ExAC, ESP, HGMD, ClinVar), *in silico* predictions (PolyPhen-2, SIFT, CADD, Mutation Taster) and ACMG classifications. After processing, variants were filtered in the determination of most likely deleterious variants using the following filtering protocol as well as variant ranking framework.

## Known gene lists

Through literature searches, tables were generated to summarize current genes known to be associated with monogenic causation of CKD. The information in these tables were used to (1) generate the curated list of known genes that would function in the exome analysis of patients and (2) to provide reference for the implication of those genes in CKD when analyzing the possible effects of those variants on the onset of patient CKD. These tables were separated into: Cystic kidney diseases (Table S.1); Proteinuric kidney diseases (Table S.2); Isolated CAKUT (Table S.3); Syndromic CAKUT (Table S.4); Tubulopathies (Table S.5); Nephritis (Table S.6); CKD and miscellaneous (Table S.7); Diabetic kidney disease (Table S.8); Hereditary amyloidosis (Table S.9). Each of these tables provides the gene name, protein products, literature reference, mode of inheritance, phenotypes, and OMIM numbers of all known monogenic causes CKD. <u>Variant filtering in WES analysis</u>

- 1. Exclude variants with allele frequencies >1% in gnomAD or ExAC databases.
- 2. Exclude synonymous and intronic variants not located within splice-site regions.

- Cross reference zygosity of variants with known disease-causing inheritance patterns and exclude variants with unmatched zygosity.
- 4. Use the gnomAD database to search for allele frequency. Exclude variants with over 10 heterozygous calls if dominant hypothesis, or 2 homozygous calls if recessive hypothesis.

#### Variant ranking in WES analysis

- Use web-based bioinformatic tools, PolyPhen-2 (polymorphism phenotyping version 2), SIFT (Sorting Intolerant From Tolerant), and MutationTaster (Berlin, Germany), and CADD (insert program used to determine this) annotated via ANNOVAR to rank the variants based on their likelihood to impact the function of the encoded protein.
- Classify mutations based on the American College of Medical Genetics and Genomics (ACMG) guidelines to predict pathogenicity of mutation, and whether the variant is pathogenic, likely pathogenic, or variant of uncertain significance (VUS)<sup>11</sup>.
- 3. For each gene selected for further analysis, conduct literature review of the gene and encoded protein, describe the known interactions, analyze structural impacts by examining variant location in protein crystal structure and structure alignment, and consult clinical histories in order to determine the likelihood of variant to cause patient phenotype.
- 4. Keep likely causative variants, and sanger confirm with segregation if possible.

#### Results

Exome sequencing was performed in a cohort of 102 people, with 94 families, 98 participants with CKD, and 4 unaffected family members. These families had undergone clinical gene panel testing, but were either undiagnosed by this tool or were unlikely to receive a diagnosis by this method, and were therefore included in this study (Fig 2).

The diagnostic rate of exome sequencing is 17% in 94 families affected by CKD who were unsolved by other means of genetic testing

Exome sequencing solved 16 familial cases of CKD, genetically, generating a diagnostic yield of (16/94) = 17% in this cohort (Fig 3). Here, a solved genetic case refers to a likely pathogenic, or pathogenic finding according to ACMG guidelines that are correlated to patient phenotypes. The causative variant annotations for this group are summarized in Table 1.

Exome sequencing had revealed 22 cases, or (22/94)=23.4% of this cohort were classified as VUS, that had strong correlations to patient phenotypes (Fig 3). These variant annotations are summarized in Table 2.

The remainder of this cohort (56/94)= 59.6% were grouped as unsolved by exome sequencing. Within this group, two patients were solved by a specific type of testing for causative variants in *MUC1*. The reason for this is variant testing for *MUC1* is not routinely detected by ES and a highly specialized type of long-range PCR is required to detect the known pathogenic variants which are located within a variable number of tandem repeat regions on the *MUC1* gene. One family had a causative variant for familial nephrolithiasis as an incidental but no variants in a gene known to cause CKD was detected. This is therefore classified as an incidental finding for this family since although it did not match the phenotype suspected as initial presentation, on review this family did indeed have a dual diagnosis of nephrolithiasis. This group, both unsolved and those with VUS findings, continue to be evaluated through familial investigations, novel gene evaluations, review of variant classifications, or re-analyses (Fig 3).

#### The prevalence of genetic kidney disease varied according to etiology in 94 families

Within genetically solved cases of CKD, underlying etiologies, prior to exome sequencing, and after this analysis, were summarized (Fig 4). These results show that a genetic basis for CKD may be variable according to specific etiologies (Fig 4). Among those solved, patients were determined to have the following etiologies, considering their molecular diagnosis: Nephronophthisis, Tubulopathy, Glomerulonephritis, Congenital anomalies of the kidneys and urinary tract (CAKUT), Tubulointerstitial kidney disease (TKD), Cystic kidney disease, and other. Within the other category, includes patients with Atypical hemolytic uremic syndrome (aHUS), Alport syndrome, and Diabetes. Importantly, prior to this analysis, 58% of patients were diagnosed with CKD of unknown etiologies, but every patient that has been solved genetically now received a specific clinical diagnosis (Fig 4).

#### Molecular diagnosis for CKD had clinical implications for patients and families

Patients who received a molecular diagnosis through exome sequencing, among which received clinical confirmation (n=13), were informed of the genetic basis of their CKD, which led to clinical implications of these findings. Among possible clinical implications, the ones that were recorded included whether the finding led to: diagnostic confirmation, resolution of diagnostic confusion, correction of diagnosis, guidance on family planning, guidance on management in transplant setting, guidance on management for living related donors, genetic counseling, cascade testing in family members, and treatment alterations. Such clinical implications were summarized and quantified according to their percent distribution in this group (Fig 5). Every solved and clinically confirmed patient had some form of clinical implication, where all cases led to diagnostic confirmation and genetic counseling (Fig 5). Following this,

among the most prevalent were diagnostic resolution/correction, while among the least prevalent was effects in the transplant and treatment settings (Fig 5).

# Case study: Dent Disease

A 48 year-old male of Italian descent was referred to the Genetic Clinic at Victoria Hospital (P101, Table1). The suspicion of a genetic basis for his CKD was raised due to early onset of CKD and ESRD as well as a positive family history of CKD. He initially presented with symptoms in his early 30s and was diagnosed with CKD at the age of 35. He recalls feeling unwell for a few weeks and after conducting blood work was immediately referred to a nephrologist. He recalls having proteinuria and advanced kidney dysfunction, however, was unsure of other specific details pertaining to his initial diagnosis. Given the small size of his kidneys, he was not a candidate for a kidney biopsy. The patient did not present with extrarenal features, but had a medical history of hypertension prior to a parathyroidectomy, steatosis, and an unprovoked pulmonary embolism. The patient received a renal transplant at the age of 46, after being on dialysis since 35 when he had reached ESRD. The patient has a positive family history of CKD, with a brother who has had a renal transplant, being diagnosed with CKD presumed due to Focal Segmental Glomerulosclerosis (FSGS) in his 40s.

Given the suspicion of a genetic basis for this patient's CKD, with negative FSGS and Alport panels, the patient was advised to receive ES. Upon sequencing, a likely pathogenic variant was found in the *CLCN5* gene (Table 1). The patient had a Guanine to Thymine mutation at position 1723, leading to a truncating variant in this gene, at position 575, on the X chromosome (Table 1). The bioinformatic annotations received through ES for this variant were a CADD score of 43 and disease causing automatic in Mutation Taster (Table 1). Given the

genetic strength of this variant, in addition to the ACMG classification, this variant was further investigated.

Inactivating mutations in the CLCN5 genes are responsible for Dent disease 1<sup>12</sup>. CLCN5 encodes a member of the voltage gated chloride ion channel family (CLC-5)<sup>13</sup>. Specifically, it encodes the 746 amino acid protein (CLC-5)<sup>13</sup>. CLC-5 proteins contain homodimers which cross the membrane in opposite directions, although each of those subunits selectively transport hydrogen and chloride ions<sup>14</sup>. The distribution of CLC-5 is significant in that it is mainly expressed in the proximal tubule and collecting ducts of the kidney, but mainly located in early endosomes<sup>16</sup>. Endocytosis through endosome acidification is essential in mediating the resorption process within the proximal tubules<sup>13</sup>. When there is a lack of functional CLC-5, the inhibition of endocytosis impacts the capacity for resorption by the proximal tubules, since it has been shown to play a vital role in this process by driving protons into cells<sup>15</sup>. Many molecules are therefore poorly resorbed<sup>12</sup>. Over 200 mutations in CLCN5, including nonsense variants, deletion variants, missense variants, frameshift variants, and intronic splicing variants have been identified to be causative of Dent disease <sup>15</sup>. Dent disease is a renal tubular disorder categorized by gene mutations in either CLCN5 or OCRL genes, into Dent Disease 1 and 2, respectively, although diseases may be phenotypically similar<sup>15</sup>.

Patients with Dent disease 1 may present with low-molecular weight proteinuria, hypercalciuria, renal calcification, nephrolithiasis, and renal tubular dysfunction<sup>16</sup>. Many patients with Dent disease develop ESRD between the ages 30-50<sup>16</sup>.

This mutation has led to the deletion of 172 amino acids at the C-terminus of *CLCN5* (Fig 6). A previous study by Lioyd et al., has shown that truncating variants, even when located upstream of this patient's mutation, leads to the abolishment of its functionality in Xenopus

oocytes<sup>17</sup>. They hypothesized that the protein's C-terminal cytoplasmic domain is essential to its function<sup>17</sup>. Within this C-terminal region, are two known cystathionine-beta synthase (CBS) domains (Fig 7), which have important regulatory function, and have been proposed to play an important role for the trafficking of this protein to the membrane where it is functional<sup>18</sup>. Additionally, previous case reports have reported a pathogenic mutation proximal to that of this patient, in a patient with Dent Disease<sup>19</sup>. Given the functional, population, and genetic data gathered regarding this variant, it was predicted to be causative of this patient's CKD, and was clinically confirmed thereafter.

This genetic diagnosis had clinical impacts for this patient. A genetic diagnosis provided a definitive clinical diagnosis for this patient, where he was previously categorized as having CKDu, and now having Dent Disease 1. He was provided with genetic counseling to explain the implications of this finding. This further prompted cascade testing in at-risk family members, and it was then established that the patient's daughter is a carrier for the condition. The clinical implication was firstly to assist in correcting the diagnosis of this patient's CKD, he was also provided with genetic counseling explaining the impacts of such a diagnosis, and provided assistance in family planning. Further, this diagnosis was able to provide cascade genetic testing in family members. The patient's daughter is now known to be a carrier for this disease, which can have impacts on her decisions in family planning. There have also been implications within the transplant setting. The patient was unaware of his diagnosis at the time of his transplant, and was therefore unable to receive adequate information regarding the possible recurrence rate of the disease. Knowing that his CKD was secondary to Dent disease, could have provided him with the knowledge regarding the recurrence of his CKD following transplant, which can vary by

etiology. Further, if living donors were being considered for transplant, assessing their risk of CKD would have been simplified with the knowledge of this diagnosis.

## Discussion

In this study, we have evaluated the utility of ES among a cohort of Canadian families that were referred to the genetic clinic due to a suspicion of having GKD, but did not receive a genetic diagnosis by routine methods. In doing so, we have calculated a diagnostic yield of 17% with clinical implications among those solved.

Studies have demonstrated that ~30% of CKD cases have underlying monogenic causation in young-onset CKD<sup>5</sup>, with evidence of a similar prevalence among adult-onset CKD<sup>7</sup> demonstrating significant genetic basis for CKD and searching for a molecular diagnosis among both cohorts of CKD patients. The diagnostic yield of ES within CKD patients has shown variability among studies, ranging from 10-50%<sup>9,10</sup>. Previous studies by our group have found a 38% yield in an Irish cohort. Here, we have shown a diagnostic yield of 17%. While this falls within the expected range, it has not shown to be as effective as other investigations through this study. However, the significant limitation in this study compared to other investigations is that we are evaluating the added benefit of ES, as opposed to its diagnostic capacity. To do this, we have only included families that remained undiagnosed by clinical panel investigations. Therefore, these families would have remained undiagnosed by current routine practice, in the absence of this tool.

Using ES, a clinical diagnosis was established for all solved families. Previous studies by our group have supported the etiological distributions that we have found here, with significant prevalence of patients with cystic kidney disease, CAKUT, and glomerulonephritis phenotypes in an adult cohort<sup>20</sup>. Within this group, 58% of patients had a clinical diagnosis of CKDu. Although

the distribution of etiologies are summarized (Fig 4), this study is not indicative of the specific genetic prevalence in subtypes of CKD, but rather representative of ES benefit in establishing a clinical diagnosis. Importantly, ES was able to provide a specific clinical diagnosis in all solved cases. Providing a patient with CKD with an accurate and specific etiology is crucial to facilitate treatment personalization in addition to establishing the prognosis of disease<sup>5</sup>.

In literature, there is significant support for a genetic diagnosis in the pursuit of personalized medicine, as it can conclude effective diagnostic procedures, aids in avoiding unnecessary procedures and treatments, helps with concluding accurate prognosis and follow-up intervals, and allows for genetic counseling for families, as well as risk calculation in the transplant setting, according to recurrence rates as well as living-donor assessments <sup>21</sup>. Here, we have assessed the specific clinical implications of a genetic diagnosis among solved families who remained undiagnosed prior to ES (Fig 5). We have shown that some form of clinical implications occurred in all families, where diagnosis confirmation, confusion resolution, correction, as well as genetic counseling were among the most prevalent (Fig 5). Nevertheless, guidance was provided in the transplant setting and with living related donors, in addition to cascade testing in at-risk family members, in some cases (Fig 5). We have shown through quantification of clinical impacts, that given the variability in prognosis, treatment, and transplant success, both among different etiologies, and within etiologies bearing different mutations, having an accurate molecular diagnosis supports the capacity to effectively treat patients with CKD.

Although there is a significant and growing association of genetic basis of many CKD cases, there are many barriers to genetic testing and hesitation to determine a genetic diagnosis. Rasouley et al., conducted a national survey among nephrologists, and showed that although all

201 nephrologists were treating GKD, 37% of these physicians only reported less than 5 patients for genetic testing<sup>22</sup>. They also demonstrated that a third of these physicians had insufficient knowledge of the genetic basis of CKD <sup>22</sup>. In literature, factors leading to hesitation for genetic testing among CKD patients include a lack of sufficient awareness for inherited diseases, accessibility to genetic testing, as well as concerns for costs <sup>23,24,25</sup>. In addition, among routine genetic testing for CKD patients, phenotype-driven gene panels are used, however these are limited in their capacity to diagnose patients whose etiologies are unknown, or patients who have been misdiagnosed, given the broad manifestations of many CKD etiologies. Here, we have shown that 17/94 families were not genetically diagnosed prior to ES. 11/17 of these families had their clinical diagnosis corrected from CKDu, to a specific etiological description. We have specifically exemplified one such case, in regards to the diagnostic methodologies as well as the clinical implications, although such an analysis was performed for every patient included in this study. Given the importance of accurate and precise diagnosis to treat patients with CKD, ES has proven to be a useful tool, where currently implemented tools are failing.

Many limitations must be considered when evaluating the results of this study. Firstly, the cohort that have been evaluated here are those who have been unable to receive a genetic diagnosis via currently implemented tools, and therefore do not represent the capacity for ES to diagnose patients with CKD as a whole, nor the prevalence of genetic causation in CKD. Further, ES is limited to the genes that are being evaluated, and as such, is only as effective as the list of genes that we have selected through literature to be valuable in searching for monogenic causation of CKD. Finally, when classifying variants according to ACMG guidelines, we have used online tools that may not have considered specific conditions which could have altered their score otherwise, such as the necessity of familial testing to accurately predict score<sup>11</sup>. As such,

many VUS variants may be classified differently, leading to a differential genetic diagnosis, if each factor in its calculation were recorded and determined specifically.

In this study, we have demonstrated the utility of ES in determining a molecular and consequently clinical diagnosis among 94 families that were undiagnosed otherwise. Given its diagnostic yield and advantages, along with the impacts that such diagnoses have had clinically, it has shown to be advantageous in routine clinical practice. Future studies should investigate whether the utility of ES can be altered with differences in the cohort, gene lists, bioinformatic tools, and altered approaches to determining variant classification via ACMG. If studies can investigate more effective methods of variant ranking/classification, as well as its utility in a broader cohort, then it can establish the importance of ES implementation in routine clinical practice, to provide a genetic diagnosis for CKD patients.

## References

- Inker, L. A., Astor, B. C., Fox, C. H., Isakova, T., Lash, J. P., Peralta, C. A., Kurella Tamura, M., & Feldman, H. I. (2014). KDOQI US commentary on the 2012 KDIGO clinical practice guideline for the evaluation and management of CKD. *American journal* of kidney diseases : the official journal of the National Kidney Foundation, 63(5), 713–735. <u>https://doi.org/10.1053/j.ajkd.20</u>
- Vaidya, S. R., & Aeddula, N. R. (2022). Chronic Renal Failure. In *StatPearls*. StatPearls Publishing. http://www.ncbi.nlm.nih.gov/books/NBK535404/
- Hill, N. R., Fatoba, S. T., Oke, J. L., Hirst, J. A., O'Callaghan, C. A., Lasserson, D. S., & Hobbs, F. D. R. (2016). Global Prevalence of Chronic Kidney Disease—A Systematic Review and Meta-Analysis. *PloS One*, *11*(7), e0158765. <u>https://doi.org/10.1371/journal.pone.0158765</u>
- 4. Arora, P., Vasa, P., Brenner, D., Iglar, K., McFarlane, P., Morrison, H., & Badawi, A. (2013). Prevalence estimates of chronic kidney disease in Canada: Results of a nationally representative survey. *CMAJ: Canadian Medical Association Journal = Journal de l'Association Medicale Canadienne*, 185(9), E417-423. <u>https://doi.org/10.1503/cmaj.120833</u>
- Connaughton, D. M., & Hildebrandt, F. (2020). Personalized medicine in chronic kidney disease by detection of monogenic mutations. *Nephrology, Dialysis, Transplantation: Official Publication of the European Dialysis and Transplant Association - European Renal Association*, 35(3), 390–397. https://doi.org/10.1093/ndt/gfz028
- 6. Neild, G. H. (2010). Primary renal disease in young adults with renal failure. *Nephrology, Dialysis, Transplantation: Official Publication of the European Dialysis and Transplant*

Association - European Renal Association, 25(4), 1025–1032. https://doi.org/10.1093/ndt/gfp653

- Connaughton DM, Bukhari S, Conlon P et al. The Irish Kidney Gene Project—prevalence of family history in patients with kidney disease in Ireland. *Nephron* 2015; 130: 293–301
- Knoers, N., Antignac, C., Bergmann, C., Dahan, K., Giglio, S., Heidet, L., Lipska-Ziętkiewicz, B. S., Noris, M., Remuzzi, G., Vargas-Poussou, R., Schaefer, F., & for the ERA Working Group on Inherited Kidney Disorders (WGIKD), which is an official body of the E. (European R. A., and the Molecular Diagnostics Taskforce of the European Rare Kidney Disease Reference Network (ERKNet). (2022). Genetic testing in the diagnosis of chronic kidney disease: Recommendations for clinical practice. *Nephrology Dialysis Transplantation*, 37(2), 239–254. https://doi.org/10.1093/ndt/gfab218
- Groopman, E. E., Marasa, M., Cameron-Christie, S., Petrovski, S., Aggarwal, V. S., Milo-Rasouly, H., Li, Y., Zhang, J., Nestor, J., Krithivasan, P., Lam, W. Y., Mitrotti, A., Piva, S., Kil, B. H., Chatterjee, D., Reingold, R., Bradbury, D., DiVecchia, M., Snyder, H., ... Gharavi, A. G. (2019). Diagnostic Utility of Exome Sequencing for Kidney Disease. *New England Journal of Medicine*, 380(2), 142–151. https://doi.org/10.1056/NEJMoa1806891
- Snoek, R., van Jaarsveld, R. H., Nguyen, T. Q., Peters, E. D. J., Elferink, M. G., Ernst, R. F., Rookmaaker, M. B., Lilien, M. R., Spierings, E., Goldschmeding, R., Knoers, N. V. A. M., van der Zwaag, B., van Zuilen, A. D., & van Eerde, A. M. (2022). Genetics-first approach improves diagnostics of ESKD patients <50 years old. *Nephrology, Dialysis,*

Transplantation: Official Publication of the European Dialysis and Transplant Association - European Renal Association, 37(2), 349–357. https://doi.org/10.1093/ndt/gfaa363

- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, Grody WW, Hegde M, Lyon E, Spector E, Voelkerding K, Rehm HL; ACMG Laboratory Quality Assurance Committee. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015 May;17(5):405-24. doi: 10.1038/gim.2015.30. Epub 2015 Mar 5. PMID: 25741868; PMCID: PMC4544753.
- Ehlayel AM, Copelovitch L. Update on Dent disease. Pediatr Clin N Am. 2019;66:169–78
- Picollo A, Pusch M. Chloride/proton antiporter activity of mammalian CLC proteins ClC-4 and ClC-5. Nature. 2005;436:420–3.
- Dutzler R, Campbell EB, Cadene M, Chait BT, Mackinnon R. X-ray structure of a ClC chloride channel at 3.0 Å reveals the molecular basis of anion selectivity. Nature. 2002;415:287–94.
- Jin, YY., Huang, LM., Quan, XF. *et al.* Dent disease: classification, heterogeneity and diagnosis. *World J Pediatr* 17, 52–57 (2021).

. .

https://doi.org/10.1007/s12519-020-00357-1

- Claverie-Martín F, Ramos-Trujillo E, García-Nieto V. Dent's disease: clinical features and molecular basis. Pediatr Nephrol. 2011;26:693–704.
- Lloyd, S. E., Pearce, S. H., Fisher, S. E., Steinmeyer, K., Schwappach, B., Scheinman, S. J., Harding, B., Bolino, A., Devoto, M., Goodyer, P., Rigden, S. P., Wrong, O., Jentsch, T.

J., Craig, I. W., & Thakker, R. V. (1996). A common molecular basis for three inherited kidney stone diseases. Nature, 379(6564), 445–449. <u>https://doi.org/10.1038/379445a0</u>

- Carr, G., Simmons, N., & Sayer, J. (2003). A role for CBS domain 2 in trafficking of chloride channel CLC-5. *Biochemical and biophysical research communications*, *310*(2), 600–605. <u>https://doi.org/10.1016/j.bbrc.2003.09.057</u>
- Okamoto, T., Tajima, T., Hirayama, T., & Sasaki, S. (2012). A patient with Dent disease and features of Bartter syndrome caused by a novel mutation of CLCN5. European journal of pediatrics, 171(2), 401–404. <u>https://doi.org/10.1007/s00431-011-1578-3</u>
- Connaughton, D. M., Kennedy, C., Shril, S., Mann, N., Murray, S. L., Williams, P. A., Conlon, E., Nakayama, M., van der Ven, A. T., Ityel, H., Kause, F., Kolvenbach, C. M., Dai, R., Vivante, A., Braun, D. A., Schneider, R., Kitzler, T. M., Moloney, B., Moran, C. P., Smyth, J. S., ... Hildebrandt, F. (2019). Monogenic causes of chronic kidney disease in adults. *Kidney international*, *95*(4), 914–928.
  - https://doi.org/10.1016/j.kint.2018.10.031
- Becherucci, F., Landini, S., Palazzo, V., Cirillo, L., Raglianti, V., Lugli, G., Tiberi, L., Dirupo, E., Bellelli, S., Mazzierli, T., Lomi, J., Ravaglia, F., Sansavini, G., Allinovi, M., Giannese, D., Somma, C., Spatoliatore, G., Vergani, D., Artuso, R., Rosati, A., ... Romagnani, P. (2023). A Clinical Workflow for Cost-Saving High-Rate Diagnosis of Genetic Kidney Diseases. *Journal of the American Society of Nephrology : JASN*, 10.1681/ASN.00000000000076. Advance online publication.

https://doi.org/10.1681/ASN.0000000000000076

22. Milo Rasouly, H., Balderes, O., Marasa, M., Fernandez, H., Lipton, M., Lin, F., Gharavi,A. G., & Sabatello, M. (2023). The impact of genetic education on referral of patients to

genetic evaluation: Findings from a national survey of nephrologists. *Genetics in medicine : official journal of the American College of Medical Genetics*, 100814. Advance online publication. https://doi.org/10.1016/j.gim.2023.100814

- 23. Curry, M., Cruz, R., Belter, L., Schroth, M., Lenz, M., & Jarecki, J. (2021). Awareness screening and referral patterns among pediatricians in the United States related to early clinical features of spinal muscular atrophy (SMA). *BMC pediatrics*, 21(1), 236. https://doi.org/10.1186/s12887-021-02692-2
- 24. Morrow, A., Chan, P., Tucker, K.M. *et al.* The design, implementation, and effectiveness of intervention strategies aimed at improving genetic referral practices: a systematic review of the literature. *Genet Med* 23, 2239–2249 (2021). https://doi.org/10.1038/s41436-021-01272-0
- 25. White, S., Jacobs, C., & Phillips, J. (2020). Mainstreaming genetics and genomics: a systematic review of the barriers and facilitators for nurses and physicians in secondary and tertiary care. *Genetics in medicine : official journal of the American College of Medical Genetics*, 22(7), 1149–1155. https://doi.org/10.1038/s41436-020-0785-6

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#### **Figures/tables**



**Figure 1. Diagnostic workflow of families being evaluated for a genetic cause to their CKD using exome sequencing as an additional tool.** Families that were referred to the genetic kidney disease clinic on suspicion of having a genetic cause to their kidney disease. This study included families that remained undiagnosed following routine clinical gene panel analysis, to evaluate the added benefit of exome sequencing in these cases. After evaluation, families were sorted into whether they were solved, whether they had a clinically significant variant of uncertain significance (VUS) and whether they were negative. Those that were solved had clinical implications, and those with VUS or negative testing continue to be evaluated by other means.

Variables	<u>Subjects</u>
Cohort size (n)	102
Unaffected family members (n)	4
Families (n)	94
CKD patients (n)	98
ESRD (n)	39
<i>Hemodialysis (n)</i>	23
<i>Renal transplant (n)</i>	16
Mean age (years)	47.5
Mean age of CKD onset (years)	34.3
Mean age of ESRD onset (years)	38.1
Ethnicity, n(%) <i>Caucasian</i> <i>Asian</i> <i>Middle Eastern</i> <i>Black or African Canadian</i> <i>Indigenous</i> <i>Hispanic</i> <i>Unknown</i>	52(51.0) 7(6.9) 2(2.0) 1(1.0) 1(1.0) 3(2.9) 36(37.2)

Variables	Subjects
CKD Etiology, n(%) Unknown Known	52(53.1) 46(46.9)
Presumed CKD Etiology, n(%) Tubulopathy Tubulointerstitial kidney disease Cystic kidney disease Glomerulonephritis CAKUT Diabetic nephropathy Other	2(2.0) 3(3.1) 9(9.2) 12(12.2) 8(8.2) 3(3.1) 9(9.2)

**Figure 2.** Details of a Canadian cohort being assessed for chronic kidney disease by exome sequencing who were previously undiagnosed. A group of n=102 people, including 98 chronic kidney disease patients undergoing exome sequencing analysis were described by being a patient or unaffected family member, whether they have reached end stage renal disease (ESRD), ESRD modality, mean current age and age of onset, etiological descriptions, and ethnicities.

CKD: chronic kidney disease; CKD etiology unknown: at the time of referral to genetic kidney disease clinic, cause of CKD was unclear; CAKUT; congenital anomalies of the kidney and urinary tracts



**Figure 3.** Exome sequencing diagnostic yield in n=94 families undiagnosed by clinical gene panel evaluations. In a group of 94 families that were unable to receive a genetic diagnosis by phenotype-targeted clinical gene panel, exome sequencing analysis was performed and its diagnostic yield was quantified. Solved is defined as having a pathogenic or likely pathogenic variant according to ACMG

guidelines that corresponds to the participant's phenotype. A variant of uncertainty (VUS) diagnosis is one that has an unknown risk associated with disease causation but is clinically relevant. Unsolved are families that were not solved genetically by this analysis.



**Figure 4. Etiological distribution of CKD patients that were genetically solved by exome sequencing analysis (n=16).** In a group of 94 families that were unable to receive a genetic diagnosis by phenotype-targeted clinical gene panel, exome sequencing analysis was performed. For genetically solved

cases (n=16), etiological distributions of CKD were summarized prior to, and following, exome sequencing analysis. In the "other" group, includes patients with Alport syndrome, Diabetes, and Atypical hemolytic uremic syndrome



**Figure 5. Prevalence of clinical implications in clinically confirmed genetic diagnoses by exome sequencing n=13.** In a group of 94 families that were unable to receive a genetic diagnosis by

phenotype-targeted clinical gene panel, exome sequencing analysis was performed. For genetically solved and clinically confirmed cases (n=13) clinical outcomes were recorded and % distribution were summarized.



# Figure 6. Representation of a nonsense mutation at position 575 of the *CLCN5* gene product.

Voltage-gated chloride ion channel 5 (CIC-5) with a premature stop codon at position 575 is shown, where the protein is coloured purple, and the deleted region is coloured in red. The specific mutation location is shown with an arrow. This figure was generated using Pymol, with PDB file P5179.



**Figure 6. Representation of a nonsense mutation at position 575 of** *CLCN5* **and the effects on known domain regions of its gene product.** Voltage-gated chloride ion channel 5 (CIC-5) with a premature stop codon at position 575 is shown, with known domain regions demonstrated. Its transmembrane, catalytic (Voltage\_CLC domain) and cystathionine beta synthase (CBS) domain are shown, where the mutation causes deletion to the C-terminal end of the catalytic domain along with both CBS domains. This figure was adapted from ENSEMBL, using the transcript number NM\_001127898.4 (ENST00000376088.7)

			<u> </u>			<u> </u>	<u> </u>												
Patient #	Family #	Gene	hg19 Pos	Transc ript	c.Change	Exon	Zygos ity	p.Cha nge	CAD D	Pol y2	SI FT	Mut Tast er	A C M G	SN P ID	Alle le Freq	ExA C	gno mA D	Cou nt	Co ver
P0071	F0048	CFH	chr1: 19664 6682	NM_0 00186	C504G	5	Het	Y168 X	23.7	//	//	А	LP	//	//	//	//	12	27
P0012	F0009	COL4 A3	chr2: 22814 2227	NM_0 00091	G2083A	28	Het	G695 R	25	D	D	D	LP	rs2 002 879 52	//	0.00 02	0.0 002	6	16
P00089	F0058	NPHP 1	2q13	//	127 kb deletion	//	Hom	//	//	//	//	//	//	//	//	//	//	//	//
P0075	F0051	UMO D	chr16: 20360 421	NM_0 010083 89	G202A	3	Het	E68K	27.9	D	D	D	LP	//	//	//	//	18	44
P0090	F0059	NPHP 1	//	//	Deletion	//	Het	//	//	//	//	//	Р	//	//	//	//	//	//
P0115	F0078	COL4 A3	chr2: 22814 2227	NM_0 00091	G2083A	28	Het	G695 R	25	D	D	D	LP	rs2 002 879 52	//	0.00 02	0.0 002	6	12
P00117	F0080	NPHP 1	//	//	106.4kb deletion	//	Hom	//	//	//	//	//	Р	//	//	//	//	//	//
P0091	F0060	PKD1	chr16: 21617 29	NM_0 00296	3437_34 39del	15	Het	1146_ 1147d el	//	//	//	//	LP	rs1 320 867 301	//	//	//	12	29
P0128	F0049	UMO D	chr16: 20360 057	NM_0 03361. 4	A665G	3	Het	Y222 C	21.2	Т	D	Ν	LP	rs1 159 216 039	//	//	//	44	89
P0140	F0097	CLC N5	chrx:4 98452 66	NM_0 00084	410dupT	5	Het	I137fs	//	//	//	//	Р	//	//	//	//	14	14
P0144	F0099	PAX2	chr10: 10253 9327	NM_0 00278	483delT	4	Het	P161f s	//	//	//	//	Р	//	//	//	//	10	23
P0101	F0067	CLC N5	chrx:4 98549 61	NM_0 00084	G1723T	10	Het	E575 X	43	//	//	А	LP	//	//	//	//	18	18
P0136	F0095	ORAI 1	chr12: 12206 4779	NM_0 32790	132delA	31	Hom	P44fs	//	//	//	//	LP	//	//	//	//	16	16
P0064	F0047	POD XL	chr7:1 31191 443	NM_0 05397	C1048T	5	Het	R382 X	35	//	//	Α	LP	rs1 351 659 066	0/1/ 313 86	//	3.2 3E- 05	25	42

Table 1: Genetic annotations of variants found to be causative of CKD in n=17 families that were solved through exome sequencing analysis.

P0183	F0125	CDK N1C	chr11: 29062 05	NM_0 00076	514_515i nsG	1	Het	P172f s	//	//	//	//	LP	//	//	//	//	6	10
P0231	F0166	CEP1 64	chr11: 11728 2575	NM_0 012719 33	c.C4213 T	31	Het	p.Q14 05X	40	//	//	Α	Р	rs1 473 989 04	0/23 0/28 237 4	0.00 08	0.0 008	31	64

Shown are patient numbers, family numbers, gene name, position on the human reference genome (Hg19pos), transcript number, nucleotide change (c.change), exon number, zygosity of the variant, amino acid change indicated by one-letter code (p.change), bioinformatic prediction tools (CADD, Poly2, Mutationtaster), ACMG classification, SNP identification, allele frequencies as per GnomAD, ExAC and GnomAD frequencies, count, and coverage of the sequencing. Abbreviations: chr=chromosome; del=deletion; ins=insertion; fs=frameshift; [bioinformatic scoring: D=deleterious; T=tolerated; A=disease causing automatic; N=polymorphism]; X=nonsense mutation; [ACMG: P=pathogenic, LP=likely pathogenic].

Table 2: Genetic annotations of clinically significant variants of unknown significance in n=22 families with CKD that were analyzed by exome sequencing

Patient #	Family #	Gene	hg19 Pos	Transc ript	c.Change	Exon	Zygos ity	p.Chan ge	CA DD	Pol y2	SI FT	Mut Tast er	A C M G	SN P ID	All ele Fre q.	ExA C	gno mA D	Coun t	Co ve r
P0015	F0012	JAG1	chr20: 10639 368	NM_0 00214	C442T	4	Het	P148S	10. 73	В	Т	D	V US	rs1 355 919 795	//	//	//	10	16
P0025	F0020	ARHG AP24	chr4: 86916 056	NM_0 01346 093	C670G	6	Het	P224A	22. 8	В	Т	D	//	rs3 552 169 5	//	0.011 3	0.00 99	23	47
P0026	F0021	REN	chr1: 20412 8683	NM_0 00537	C533T	5	Het	T178M	26. 3	D	Т	D	V US	rs1 474 368 51	//	6.59 E-05	3.23 E-0 5	9	35
P0088	F0057	FN1	chr2:2 16279 564	NM_0 01306 129	G1937A	13	Het	R646K	15. 89	В	Т	D	V US	//	//	//	//	17	31
P0103	F0068	CFI	chr4: 11068 5820	NM_0 00204	G355A	3	Het	G119R	22. 3	D	D	Ν	V US	rs1 418 535 78	//	0.000 5	0.00 05	18	40
P0099	F0065	UMOD	chr16: 20360 277	NM_0 01008 389:	G346C	3	Het	G116R	23. 5	D	D	Ν	V US	rs1 390 427 57	//	//	//	33	70
P0200	F0140	TTC21 B	chr2: 16678 6721	NM_0 24753	G1048T	9	Het	A350S	28. 6	D	Т	D	V US	//	0/1 6/2 510 82	7.42 E-05	//	13	28
P202	F0141	COL4 A5	Chr2: 22817 2594	NM_0 00091	T4421C	48	Het	L1474 P	23. 4	D	D	D	V US	//	//	0.002 8	0.00 22	13	30

P202	F0141	KCNJ1 1	Chr11: 17409 548	NM_0 00522	C91T	1	Het	R31W	//	//	//	//	V US	//	0	//	//	29	60
P0185	F0127	TBC1D 8B	chrX: 10611 7191	NM_0 17752	c.T3359G	21	Het	p.M11 20R	17. 24	В	D	Ν	V US	//	//	//	//	16	16
P0028	F0023	APOE	chr19: 45409 886	NM_0 00041	A5G	2	Het	K2R	20. 7	В	Т	N	V US	//	//	//	//	18	41
P0021	F0016	CYP11 B1	chr8: 14395 8612	NM_0 00497	G422A	3	Het	R141Q	26. 8	D	D	D	V US	rs2 676 018 10	0/3/ 281 926	9.00 E-06	6.47 E-0 5	14	35
P0047	F0033	SLC12 A3	chr16: 56920 296	NM_0 00339	C1946T	16	Het	T649M	32	D	D	D	V US	rs1 453 376 02	//	0.000 0498	0.00 006 47	3	11
P0054	F0037	KLHL3	chr5: 13696 1453	NM_0 01257 195	G1478A	12	Het	R493Q	35	D	D	D	LP	//	//	//	//	12	35
P0095	F0062	SLC6A 19	chr5:1 21067 6	NM_0 01003 841	C461T	3	Het	P154L	27. 6	D	D	D	V US	rs7 719 807 56	0/9/ 281 662	4.18 E-05	3.23 E-0 5	35	77
P0059	F0042	ATXN1	6:1632 7865	NM_0 00332. 4	677_678i nsGCA	8	Hom	Q225d up	//	//	//	//	V US	//	321 /91 5/1 008 40	//	//	//	//
P0059	F0042	ATXN1	6:1632 7916	NM_0 00332. 4	621_626d upGCAG CA	8	Het	Q207_ Q208d up	//	//	//	//	V US	//	3/6/ 182 42	//	//	//	//
P0107	F0070	LAMC 1	chr1:1 83096 481	NM_0 02293	A3065G	17	Het	N1022 S	23. 8	D	D	D	V US	rs1 477 946 01	0/4 2/2 828 70	0.000 1	9.69 E-0 5	12	28
P0109	F0072	SLC6A 19	chr5: 12125 05	NM_0 01003 841	C569T	4	Het	S190L	26. 2	Р	D	D	V US	//	//	2.478 E-05	//	38	71
P0121	F0084	NPHP3	chr3:1 32416 142	NM_1 53240. 5	G2050T	14	Hom	E684X	44	//	//	A	V US	//	//	//	//	4	12
P0135	F0094	TSC1	chr9: 13580 0991	NM_0 00368	T346G	5	Het	L116V	22. 7	D	Т	D	V US	rs1 996 202 68	0/1 13/ 282 648	0.000 3	0.00 05	21	44
P0168	F0115	GATA3	chr10: 81004 80	NM_0 01002 295	A454C	3	Het	T152P	23	Р	Т	D	V US	//	//	//	//	35	77
P0234	F0169	ABCC 6	chr16: 16259 743	NM_0 01171	c.C3043T	23	Het	p.R101 5W	26. 4	D	Т	N	V US	//	0/7/ 281 098	1.687 E-05	3.23 4E- 05	11	29

P0186	F0128	BICC1	chr10: 60546 719	NM_0 01080 512	A424G	5	Het	T142A	25. 8	D	D	D	V US	//	//	//	//	9	23
P0186	F0128	FN1	chr2: 21628 5470	NM_0 01306 129	G1601A	11	Het	R534H	35	D	D	D	V US	rs1 421 650 52	0/6 0/2 828 00	0.000 2	0.00 03	7	16

Shown are patient numbers, family numbers, gene name, position on the human reference genome (Hg19pos), transcript number, nucleotide change (c.change), exon number, zygosity of the variant, amino acid change indicated by one-letter code (p.change), bioinformatic prediction tools (CADD, Poly2, Mutationtaster), ACMG classification, SNP identification, allele frequencies as per GnomAD, ExAC and GnomAD frequencies, count, and coverage of the sequencing. Abbreviations: chr=chromosome; del=deletion; ins=insertion; fs=frameshift; dup=duplication; [bioinformatic scoring: B= benign; D=deleterious; T=tolerated; N=polymorphism]; X=nonsense mutation; [ACMG: VUS=variant of unknown significance].

## **Supplemental Items**

Supplementary Table S1: genes that represent monogenic causes of human cystic kidney disease or nephronophthisis, if mutated.

Jene	1 <i>lias</i>	<sup>•</sup> rotein	<i>leference</i>	10de nheritan	ofhenotype ce	)MIM number
1LMS1	1LMS	Iltrom Syndrome Protein	17.01lin Nat 11.11:74, 2002	Genet1R	llstrom syndrome	ŧ 203800
1LG9		1LG9 Ipha-1,2-mannosyltransf ase	Veinstein Am. J. Geret. 136A: 19 2005	Med.1R 4-197,	Congenital disorder glycosylation, type Il	ofi06941
39D1	1KS9	39 domain containin vrotein l	głomani Orphan łare Dis 9:72, 20	net JIR )14	<i>Aeckel syndrome 9,</i> <i>'oubert syndrome 27</i>	‡ 614209, ‡ 617120
39D2	<i>AKS10</i>	39 domain containin vrotein2	g)owdle AJHG <sup>1</sup> 4, 2011	89(1):1R	Aeckel syndrome oubert syndrome 34	10,‡614175
3BS1		<i>Sardet-Biedl Syndrome 1</i>	<i>Aykytyn Nat</i> 1(4):435, 2002	GenetlR	Sardet-Biedl syndrome	1 ±209900
3BS2		<i>Sardet-Biedl Syndrome 2</i>	Catsanis S 193(5538):2256,	ciencelR 2001	Sardet-Biedl syndrome	2 ±615981
3BS3	1 <i>RL6</i>	<i>Meckel syndrome, type 1</i>	<i>Chaddour Hum</i> 28(5), 523, 2007	MutatlR	Sardet-Biedl syndrome	3 ±600151
3 <i>BS4</i>		3ardet-Biedl Syndrome 4	<i>Aykytyn</i> Nat ≥(2):188, 2001	GenetlR	Sardet-Biedl syndrome	4 ±615982
3BS5		3ardet-Biedl Syndrome 5	<i>Fieder Int J F</i> <i>Vephrol 3(3):199</i>	Pediatr1R , 1982	<i>Sardet-Biedl syndrome</i>	5 ±615983
3BS6	AKKS	<i>Sardet-Biedl Syndrome 6</i>	Catsanis Nat 26(1):67, 2000	GenetlR	3ardet-Biedl syndrome	6 ±605231

3BS7		<i>Sardet-Biedl Syndrome 7</i>	3adano AJHG 72(3),1R 550, 2003	3ardet-Biedl syndrome 7 ±615984
3BS8	"TC8	<i>Sardet-Biedl Syndrome 8</i>	<i>Stoetzel J Hum Genet</i> ( <i>R</i> ) <i>1(1):81, 2005</i>	<i>Bardet-Biedl syndrome 8 ±615985</i>
3BS9	'THB1	Sardet-Biedl Syndrome 9	√ishimura AJHG1R '7(6):1021, 2005	3ardet-Biedl syndrome 9 ±615986
3BS10		<i>Sardet-Biedl Syndrome 10</i>	<i>Stoetzel Nat GenetR</i> <i>8(5):521, 2006</i>	3ardet-Biedl syndrome 10 <sup>‡</sup> 615987
3BS11	"RIM32	Sardet-Biedl Syndrome 11	Chiang PNAStR 03(16):6287,2006	3ardet-Biedl syndrome 11 <sup>4</sup> 615988
3BS12		Sardet-Biedl Syndrome 12	Stoetzel AJHG 80(1):1,1R 2007	<i>Bardet-Biedl syndrome 12</i> <sup>‡</sup> 615989
3BS15	VDPCP	VD repeat-containing vlanar cell polarit ffector	gitone Nat GenetlR v5(1):79, 2000	8ardet-Biedl syndrome 15 <sup>‡</sup> 615992
3BS17	ZTFL1	<i>Bardet-Biedl syndrome 17</i>	<i>Aarion J Med Genet</i> ( <i>R</i> 19(5):317, 2012	3ardet-Biedl syndrome 17 <sup>‡</sup> 615994
3BS18	3BIP1	<i>Sardet-Biedl syndrome 18</i>	Scheidecker J MedlR Genet 51(2):132, 2014	Bardet-Biedl syndrome 18 <sup>‡</sup> 615995
3BS20	FT27	3ardet-Biedl Syndrome 20	<i>Schaefer J Med Genet R 51(5):447 2016</i>	Bardet-Biedl syndrome 19 <sup>‡</sup> 615996
C2CD3	)FD14	Drofaciodigital syndrom 4	e <sup>-</sup> hauvin-Robinet 1R Vature Genet '6(8):905, 2014.	Drofaciodigital syndrome <sup>‡</sup> 615948 (IV
CCDC28E	}	Coiled-coil domain ontaining protein 28B	1 <sup>°</sup> ardenas-Rodriguez 1R/D Tum Genet 132(1):91, V013	DR Bardet-Biedl syndrome 1,±209900 nodifier of
CCDC41	CEP83	Coiled-coil domain ontaining 41	<i>v<sup>z</sup>ailler</i> AJHG1R <i>v</i> 4(6):905, 2014	<i>Vephronophthisis 18 ± 615862</i>
CCND1		Cyclin D1	'u The Journal oflD vrology, 172(6 Pt 1), \'410–2413, 2004	<i>Renal cell carcinoma</i> 68461
CDC73		Cell division cycle 73	<sup>°</sup> eh J. Clin. Endocr.1D 1etab. 81: 4204-4211, '996	Parathyroid adenomai07393 vith cystic changes
CEP120	RTD13	Centrosomal protein 20kDa	n <sup>3</sup> haheen Hum MollR Tenet 24(5):1410, '015	Whort-rib thoracic <sup>‡</sup> 616300, Nysplasia 13 with or± 617761 Vithout polydactyly, Noubert syndrome 31
CEP41	"SGA14	Centrosomal protein !1kDa	n.ee Nat GenetlR !4(2):193, 2012	oubert syndrome 15 4614464

COL4A1		Collagen, type IV, alpha-1	Plaisier New Eng. J.1D Aed. 357: 2687-2695, 2007	Ingiopathy, hereditary, 20130 vith nephropathy, neurysms, and muscle ramps; hematuria, renal ysts
CPT2		Carnitine	<i>Cinn Pediat. Res. 29\D/AR</i> suppl.): 73A only, 991	CPT II Deficiency 600650
)DX59	)FD5	Drofaciodigital syndrom	e'hamseldin AJHG1R V3(3):555, 2013	<b>Drofaciodigital syndrome</b> <sup>‡</sup> 174300
)YNC2HI	RTD3	<i>Dynein cystoplasmic</i> <i>eavy chain</i>	271 Hokayem J MedlR Fenet 49(4):227, 2012	Nort-rib thoracic <sup>t</sup> 613091 lysplasia 3 with or vithout polydactyly
FANI	<i>4TMR15</i>	<i>ANCI-associated</i> <i>nuclease 1</i>	Thou     Nat     GenetIR       14(8):910, 2012     14	nterstitial nephritis, <sup>‡</sup> 614817 aryomegalic
<i>JANAB</i>		Flucosidase, alpha weutral AB	1, <sup>2</sup> orath Am. J. Hum.1D Fenet. 98: 1193-1207, 2016	Polycystic kidney disease 04160
7 <i>LIS3</i>		<i>3LIS family zinc finge</i> vrotein 3	r <sup>r</sup> aha Am. J. Med.1R Fenet. 122A: 269-273, '003	Diabetes mellitus, 10192 weonatal, with congenital hypothyroidism
HOXA4		Teomobox A4	tcampora     NucleicID       tcids     Res       '7(24):10385, 1985	VA <sup>•</sup> 142953
НОХВ6		Heomobox B6	Caur J Exp ZoollD 164(3):323, 1992	VA *142961
HSD17B4		7-Beta-Hydroxysteroid lehydrogenase IV	<sup>7</sup> erdinandusse Am. J.1R Ium. Genet. 78: 12-124, 2006	D-bifunctional protein <sup>(01860</sup> leficiency; renal cysts, drenal cortex atrophy
FT43	CED3	ntraflagellar transport 43	3 <i>Filissen AJHG</i> 1R 37(3):418, 2010	Cranioectodermal lysplasia 3 ± 614099
FT52	RTD	ntraflagellar Transport 5.	27irisha Clin GenetlR V0(6):536, 2016	Whort-rib thoracic <sup>t</sup> 617102 Pysplasia 16 with or vithout polydactyly
FT57		ntraflagellar Transport 5	73ruel J Med GenetlR :4(6):371, 2017	)rofaciodigital syndrome <sup>t</sup> 617927 (VIII
FT80	SRTD2	ntraflagellar Transport 8	03eales Nat GenetlR 19(6):727 2007	<i>Whort-rib thoracic</i> <sup>£</sup> 611263 <i>Sysplasia 2 with or vithout polydactyly</i>
FT81	CDV-1	ntraflagellar Transport 8	Perrault J Med GenetlR 52(10):657, 2015	Nort-rib thoracic 605489 lysplasia 19 with or vithout polydactyly

FT122	CED1	ntraflagellar transpo 22	rtValczak-Sztulpa AJHG1R 36(6):949, 2010	Cranioectodermal lysplasia 1	±218330
<i>FT140</i>	RTD9	ntraflagellar transpo '40	rPerrault AJHGIR 90(5):864, 2012 Senum AJHG 109(1):1D 536-135	Short-rib thora lysplasia 9 with vithout polydactyly vutosomal domine volycystic idney-spectrum	cic <sup>‡</sup> 266920 or ant <sub>Va</sub>
BTS1	NPP5E	nositol volyphosphate-5-phospha ise	3ielas Nat GenetlR ht !1(9):1032, 2009	'oubert syndrome 1	<i>±213300</i>
BTS2	"MEM21	6'ransmembrane Protei '16	inEdvardson AJHGIR 26(1):93, 2010	Aeckel syndrome 2, 'oubert syndrome 2	‡ 603194 ‡ 608091
BTS3	1 <i>HI1</i>	1belsonHelpentegration Site 1	er <sup>3</sup> arisi J Med GentR !3(4):334 ,2005	oubert syndrome 3	±608629
BTS8	1 <i>RL13B</i>	1DP-ribosylation `actot-like 13B	Cantagrel AJHG1R 3(2):170, 2008	oubert syndrome 8	£ 612291
'BTS9	CC2D2A	Coiled-coil and C lomains-containing vrotein 2A	2Voor AJHG1R 32(4):1011, 2008	'oubert syndrome 9 Aeckel syndrome 6 COACH syndrome	‡ 612285, ‡ 612284, ‡ 216360
BTS12	CIF7	<i>Kinesin family member 7</i>	Putoux Nat GenetlR !3(6):601, 2011	1crocallosal syndro 'oubert syndrome 12	me‡ 200990
BTS13	"CTNI	ectonic family member 1	Garcia-Gonzalo NatlR Genet 43(8):776, 2011	oubert syndrome 13	£ 614173
BTS14	"MEM23	7 <sup>r</sup> ransmembrane protei !37	inłuang AJHGIR 39(6):713, 2011	oubert syndrome 14	£ 614424
BTS16	"MEM13	8 <sup>°</sup> ransmembrane protei '38	in.ee Science 335(6071):1R 966, 2012	oubert syndrome 16	± 614465
BTS17	25orf42	Chromosome 5 ope eading frame 42	nšrour AJHG 90(4):693,1R '012	oubert syndrome 17	± 614615
BTS18	"CTN3	<i>Tectonic family member 3</i>	Thomas     AJHGIR       91(2):372, 2012     111111111111111111111111111111111111	oubert syndrome 18	£ 614815
BTS20	"MEM23	1 <sup>°</sup> ransmembrane protei '31	inšrour J Med Genet 49:1R 536-641, 2012	'oubert syndrome 20 Aeckel syndrome 11	‡ 614970, ‡ 615397
BTS21	CSPP1	Centrosome spind vole-associated protein 1	letkizu AJHG 94(1):80,1R '014	'oubert syndrome 21	£ 615636
BTS22	'DE6D	Phosphodiesterase 6D	"homas Hum MutatlR 25(1):137, 2014	oubert syndrome 22	£ 615665

BTS23	<i>(IAA058)</i>	6"ALPID 3, chicken vomolog of	Bachmann-Gagescu Tum Mutat 36(9):83 V015	1R 31,	<i>Foubert syndrome 23</i>	616490
BTS24	"CTN2	<i>Fectonic family member 2</i>	Huppke Eur J Hu Fenet 23(5):616, 201	um1R 5	<i>'oubert syndrome 24</i>	616654
BTS25	<i>CEP104</i>	Centrosomal protein !1kDa	nkorvatska Am J Mo Fenet Veuropsychiatr Gen V56B(3):303, 2011	iedIR B net	<i>Foubert syndrome 25</i>	616781
BTS26	(IAA055)	6 Catanin-intereacting vrotein	Saunders Genome Bi 6:293, 2015	iollR	<i>Soubert syndrome 26</i>	514175
<i>(1AA0753</i>	)FD15	Drofaciodigital syndrom 5	e]hevrier Hum M Fenet 25(3):497, 201	lollR 6	Drofaciodigital syndrome <sup>t</sup> (V	617127
CIF14	<i>AKS12</i>	Kinesin family member 14	Filges Clin Gen 6(3):220, 2013	1etlR	<i>Aeckel syndrome 12</i> to	616258
AKS1		Aeckel syndrome, type 1	(yttälä Nat Ger 18(2):155, 2006	netlR	Aeckel syndrome 1#2'oubert syndrome 28#Bardet-Biedl syndrome 13#	249000, 617121, 615990
VEK1	RTD6	VIMA Related Kinase 1	"hiel AJHG 88(1):10 2011	)6,1R	whort-rib thoracict lysplasia 6 with or vithout polydactyly	263520
VPHP1		Vephrocystin 1	Hildebrandt Nat Gen 7(2):149, 1997	netlR	'uvenile nephronophthisist ' t 'oubert syndrome 4 t 'enior-Loken syndrome-1	256100 609583 266900
VPHP2	NVS	nversin	Otto Nat Gen 14(4):413, 2003	1etlR	nfantile nephronophthisis <sup>‡</sup> (	602088
VPHP3		<i>Vephrocystin 3</i>	<i>Ilbrich Nat Gen</i> 14(4):455, 2003	netlR	Vephronophthisis 3 <sup>‡</sup> ( Aeckel syndrome <sup>‡</sup> ) Renal-hepatic-pancreatic <sup>‡</sup> ) lysplasia 1	604387 267010 208540
VPHP4		Vephronophthisis 4	Dtto AJHG 71(5):116 2002	51,1R	Vephronophthisis 4 to the second seco	606966
VPHP5	QCB1	Q motif containing B1	Otto     Nat     Gen       17(3):282, 2005     17	1et1R	Senior-Loken syndrome 5 to	609254
VPHP6	<i>CEP290</i>	Centrosomal protein '90kDa	niayer Nat Gen 18(6):674, 2006	net1R	Foubert syndrome 5 #   Bardet-Biedl syndrome 14# #   Aeckel syndrome 4 #	610188 615991 610189
VPHP7	FLIS2	3LIS Family Zinc Finger	21ttanasio Nat Gen 19(8):1018, 2007	netlR	<i>Vephronophthisis 7</i>	611498

VPHP8	₹PGRIP1	<i>PGRIP1 Like</i> 1 <i>rts Nat Genet</i> 11 <i>!9(7):882, 2007</i>	RAeckel syndrome 5± 611561,'oubert syndrome 7± 611560,COACH syndrome± 216360
VPHP9	VEK8	VIMA (never in mitosis)tto JASN 19(3):587,11 gene a) - related kinase 8 '008	R Renal-hepatic-pancreatic ± 615415 lysplasia 2 ± 613824 Vephronophthisis 9
VPHP10	DCCAG	&erologically Defined)tto Nat Genet(1 Colon Cancer Antigen 8 !2(10):840, 2010	R 3ardet-Biedl syndrome 16±615993 Senior-Loken syndrome 7 ±613615
VPHP11	"MEM67	'ransmembrane     Protein)tto     J     Med     Genet11       57     !6(10):663, 2009	R COACH syndrome, <sup>‡</sup> 216360 Vephronophthisis 11 ±613550
VPHP12	"TC21B	"etratricopeptideRepeatDavisNatGenetIlDomain 21B!3(3):189, 2011	R Vephronophthisis 12 ± 613820
VPHP13	VDR19	VD repeat domain 19     3edrup     AJHGII       39(5):634, 2011     39(5):634, 2011     39(5):634, 2011	R Vephronophthisis 13 ± 614377 Senior-Loken syndrome 8 ± 616307
VPHP14	ZNF423	<i>Linc finger protein 423 Chaki Cell 150(3):533,11</i> 2012	R Vephronophthisis 14 ± 614844 Soubert syndrome 19
VPHP15	CEP164	Centrosomal proteinChaki Cell 150(3):533,11 '64kDa '012	R Vephronophthisis 15 ±614845
VPHP16	1NKS6	Inkyrin repeat and sterileIoffNatGenetIIslphamotifdomain!5(8):951, 2013ontaining 6	R Vephronophthisis 16 ±615382
VPHP17	FT172	ntraflagellar transportIalbritter AJHGI 72 homolog, 3(5):915, 2013 Chlamydomonas	R Whort-rib thoracic <sup>t</sup> 615630 lysplasia 10 with or vithout polydactyly
VPHP19	)CDC2	Double-cortindomain chuelerAJHGIrontaining protein 206(1):81, 2015	R Vephronophthisis 19 ±616217
VPHP20	/APKBP	Aitogen activated protein/aciaAJHGIinase-binding protein 1'00(2):323, 2017	R Jephronophthisis 20 ± 617271
VPHPL1	(PNPEP3	C-prolyl aminopeptidase 3 D'Toole J Clin Investil 20(3):791, 2010	R Vephronophthisis-like ±613159 vephropathy 1
°KHD1	1RPKD	PKHD1,Bergmann Kidney IntllFibrocystin/Polyductin57(3):829 2005	<i>Polycystic kidney disease</i> <sup>£</sup> 263200 <sup>!</sup> with or without hepatic <sup>!</sup> isease
°OC1B		Cone rod dystrophy 20 Roosing AJHG11 15(2):131, 2014	Cone-rod dystrophy 20
CLTI	)FD9	Drofaciodigital syndromeldly Hum Mutation11))<	R Drofaciodigital syndrome%258865 X
LC41A1		<i>Solute carrier memberIurd JASN 24(6):967,11</i> <i>amily 41, member 1</i> 2013	R VA <sup>°</sup> 610801

"MEM107 4KS13	Transmembrane Proteinihasheen Hum MollR 107 Fenet 24(18):5211, 1015	<i>Aeckel syndrome 13 4617562</i> <i>'oubert syndrome 29</i>
"RAF3IP1 \LS9	"NF receptor-associatedBerbari Dev BiollR "actor 3-interacting'60(1):66, 2011 vrotein 1	Senior-Loken syndrome 9 ± 616629
JSH2A	Jsherin 2A Smith GenomicslR 4(4):995, 1992	Jsher syndrome type 2A ±276901
VDR34 \RTD11	VD Repeat Domain 34 Schmidts AJHG1R 93(5):932, 2013	Nort-rib thoracic <sup>‡</sup> 615633 lysplasia 11 with or vithout polydactyly
VDR35 CED2	VD repeat domain 35 Filissen AJHG1R 37(3):418, 2010	Cranioectodermal ±613610 lysplasia 2
VDR60 \RTD8	VD Repeat Domain 60 <i>AcInerney-Leo AJHG</i> IR V3(3): 515, 2013	Nort-rib thoracic <sup>‡</sup> 615503 lysplasia 8 with or vithout polydactyly
1LG5	1LG5 Dolichyl-Phosphate.emoine Am J Hum1D 3eta-Glucosyltransferase Jenet 109:1484, 2022	Polycystic Kidney <sup>‡</sup> 620056 Disease 7
RP5	Low density lipoprotein Inossen proc NatllD eceptor-related protein 5 lcad Sci U S A 11:5343, 2014	Polycystic liver disease 4 <sup>‡</sup> 617875 vith or without kidney lisease
ABTPS2	Membrane-boundYeligmanArch1DranscriptionFactor)ermatol 80:413, 1959Protease, Site 2	FAP Syndrome 1, with or <sup>‡</sup> 308205 vithout Bresheck <sup>3</sup> yndrome
EC63	EC63 Homolog, ProteinInossen Orphant JlD Franslocation regulator Care Dis 1:69, 2014	<i>Polycystic liver disease 2 ± 617004</i>
PRKCSH	Protein Kinase C3errebi Clin GenetlD Substrate, 80-KD, Heavy1:342,1982 Shain	Polycystic liver disease 1 ± 174050
"BC1D32	"BC1 Domain FamilyIdly Hum Mutat 35:36,1DMember 322014	VA <sup>°</sup> 615867
<i>WC</i>	EvC CiliaryComplex?uiz-PerezNatGenet.1R/ADSubunit 124(3):283, 2000	Ellis-van Creveld <sup>‡</sup> 225500 yndrome
EVC2	EvCCiliaryComplex/urianIndianJDent/R/ADSubunit 2 <a><a><a><a><a><a><a><a><a><a><a><a><a></a></a></a></a></a></a></a></a></a></a></a></a></a>	Ellis-van Creveld <sup>‡</sup> 225500 yndrome
)FD1	)rofaciodigital syndrome Feather Hum Mol(LD Genet 6(7):1163, 1997	Drofaciodigital syndrome <sup>‡</sup> 311200

*AR*, autosomal recessive; *AD*, autosomal dominant; *DR*, digenic recessive; *NA*, not available; *OMIM*, *Online Mendelian Inheritance in Man; XLD; X-linked dominant; XL; X-linked; #, phenotype MIM number; \* gene/ locus MIM number if not phenotype MIM number available.* 

Jene	Protein	<i>leference</i>	1ode nheritance	of henotype ?	OMIM sumber
1DCK4	<i>larF domain containin</i> <i>inase 4</i>	nglshraf J Clin Invest 123:517 '013	'9,1R	<i>Vephrotic syndrom</i> <i>ype</i> 9	e,° 615567
1LG1	1LG1, Thitobiosyldiphosphodolich Beta-Mannosyltransferase	Iarshman Pediatr Int 58:78 0'016	5,1R	Congenital disorde of glycosylation typ k	er‡608540 9e
<i>IRHGDIA</i>	tho GDP dissociation hibitor (GDI) alpha	mree J Clin Invest 123:324 '013	3,1R	<i>Vephrotic syndrom</i> <i>ype</i> 8	e,‡ 615244
1VIL	1dvillin	Rao J Clin Invest 127:425 2017	7,1R	VA	° 613397
CD2AP	CD2 associated protein	Cim Science 300:1298, 2003	1R	Flomerulosclerosis, ocal segmental, 3	ŧ 607832
COQ2	Coenzyme Q !-hydroxybenzoate volyprenyltransferase	2Diomedi-Camassei JAS '8:2773, 2007	SN1R	Primary coenzym 210 deficiency 1	ne <sup>‡</sup> 607426
COQ6	Coenzyme Q vonooxygenase	964eeringa J Clin Inve '21:2013, 2011	est1R	Primary coenzym 210 deficiency 6	ae‡ 614650
CUBN	Cubilin (intrinst `actor-cobalamin receptor)	ic)vunc JASN 22:1815, 2011	1R	Aegaloblastic memia-1, Finnis ype	‡261100 :h
CRB2	Crumbs, Drosphilia Homolog of 2	a,Ebarasi AJHG 96: 153-16 '015	1,1R	Focal segmenta clomerulosclerosis S	al# 616220 )
)GKE	Diacylglycerol kinase epsilo	on.emaire Nat Genet 45: 53 '013	1,1R	<i>Vephrotic syndrom</i> <i>ype</i> 7	e,‡ 615008
EMP2	Epithelial membrane protei	iniiee AJHG 94:884, 2014	1R	<i>Vephrotic syndrom</i> <i>ype 10</i>	e,‡615861
7AT1	<sup>7</sup> at tumor suppresso Irosphila, homolog of, 1	r;7ee Nat Commun 7:10822, 201	161R	VA	° 600976
TGA3	ntegrin, alpha 3 (antige CD49C, alpha 3 subunit o 'LA-3 receptor)	en'alcin Hum Mol Genet 24:367 of 015	9,1 <i>R</i>	Congenital nterstitial lun lisease, nephrot yndrome, an pidermolysis bullos	±614748 g ic ed ca

Supplementary Table S2: genes that represent monogenic causes of human nephrotic syndrome, if mutated.

TGB4	ntegrin, beta 4	<i>Cambham AJKD 36:190, 2000</i> 1 <i>R</i>	Epidermolysis ±226730 vullosa
ZANKI	<i>IN motif and ankyrin reper- lomain-containing protein</i>	atiee J Clin Invest 125:2375,1R 1 '015	Cerebral palsy,±612900 pastic quadriplegic,
KANK2	(N motif and ankyrin repersion of the second	atree J Clin Invest 125:2375,1R 2 '015	√ephrotic syndrome,±617783 ype 16
CANK4	<i>IN motif and ankyrin reper- lomain-containing protein</i> .	atree J Clin Invest 125:2375,1R 3 '015	VA <sup>°</sup> 614612
AGE3	antigen family member 3	3raun Nat Genet 49:1529, 2017 1R	Falloway-Mowat ±301006 yndrome 2, X-linked
AMB2	aminin, beta 2	Yenker Hum Mol Genet 12:2625,1R Y004	Vephrotic syndrome,±614199 ype 5, with or vithout ocular sbnormalities
CAT	ecithin-Cholesterol Cyltransferase	<sup>r</sup> aramelli Hum Genet 85:195,1R '990	√orum disease
1AGI2	<i>Membrane-associated juanylate kinase, WW an</i> PDZ domains-containing 2	3ierzynska JASN 28:1614, 2017 IR d	<i>Vephrotic syndrome,</i> <sup>±</sup> 617609 <i>ype</i> 15
<i>ΛΥΟΙΕ</i>	Iomo sapiens myosin I MYO1E)	E1ele NEJM 365:295, 2011 1R	Flomerulosclerosis, ±614131 ocal segmental, 6
VEU1	Neuraminidase 1	<i>Aütze Genet Metab Rep 10:1-4,1R</i> '016	ialidosis ±256550
VPHS1	Vephrin	<i>Cestila Mol Cell 1:575, 1998</i> 1 <i>R</i>	Vephrotic syndrome,‡256300 ype 1
VPHS2	Podocin	<i>Boute Nat Genet 24:349, 2000</i> 1 <i>R</i>	Vephrotic syndrome,‡ 600995 ype 2
<i>VUP107</i>	Vucleoporin, 107-KD	<i>Aiyake AJHG 97:555, 2015</i> 1 <i>R</i>	√ephrotic syndrome,±616730 ype 11
VUP133	Vucleoporin 133-KD	3raun Nat Gene 48:457, 2016 1R	VA \$607613
VUP205	Jucleoporin, 205-KD	3raun Nat Gene 48:457, 2016 1R	Vephrotic syndrome, <sup>‡</sup> 616893 ype 13
VUP85	Vucleoporin 85-KD	3raun Nat Gene 48:457, 2016 1R	VA <sup>°</sup> 170285
<i>VUP93</i>	Jucleoporin, 93-KD	3raun Nat Gene 48:457, 2016 1R	√ephrotic syndrome,‡616892 ype 12
<i>SGEP</i>	D-sialoglycoprotein mdopeptidase	3raun Nat Genet 49:1529, 2017 1R	Falloway-Mowat \$617729 yndrome 3
DSS2	Prenyl (decapreny liphosphate synthas ubunit 2	l).opez AJHG 79:1125, 2006 IR e,	Primary coenzyme <sup>1</sup> 614652 210 deficiency 3

PLCE1	Phospholipase C, epsilon 1 Finkes Nat Genet 38:1397, 20061R	Jephrotic syndrome, <sup>‡</sup> 610725 ype 3
PTPRO	ProteintyrosineDzaltin AJHG 89:139, 2011IRvhosphatase, receptor type,)	Vephrotic syndrome,±614196 ype 6
CARB2	Scavenger receptor class B,3adhwar Brain 127: 2173, 2004 IR nember 2	Epilepsy, progressive <sup>1</sup> 254900 nyoclonic 4, with or vithout renal failure
GPL1	Sphingosine1phosphate.ovricJClinInvest127:912,1Rvase 12017	<i>Vephrotic syndrome,</i> <sup>‡</sup> 617575 <i>ype</i> 14
MARCALI	WI/SNF related, matrix3oerkoel Nat Genet 30:215,1R issociated, actin dependent'002 egulator if chromatin, subfamily	Schimke ‡242900 mmunoosseous Iysplasia
TP53RK	<i>"P53-regulating kinase 3 raun Nat Genet 49:1529, 2017 1R</i>	Falloway-Mowat ±617730 yndrome 4
"PRKB	<i>"P53RK binding protein 3 raun Nat Genet 49:1529, 2017 IR</i>	Falloway-Mowat ±617731 yndrome 5
TTR	TransthyretinIndoBiochemBiophysRestRCommun 211:354, 1995	Hereditary ± 105210 ransthyretin-related ymyloidosis
PS33B	<sup>7</sup> PS33B, Late Endosome <sup>3</sup> ull J Pediatr 148:269, 2006 1R 1nd Lysosome Associated	VA <sup>°</sup> 608552
VDR73	<i>VD</i> repeat-containing Colin AJHG 95:637, 2014 1R vrotein 73	Falloway-Mowat ±251300 yndrome 1
CPO5	Exportin 5 3raun Nat Genet 48:457, 2016 1R	VA <sup>607845</sup>
1CTN4	1ctinin, alpha 4Caplan Nat Genet 24(3):251,1D2000	Flomerulosclerosis, ±603278 `ocal segmental 1
INLN	1ctin-binding protein anillin 7badegesin JASN 25:1991, 20141D	Focal segmental <sup>‡</sup> 616032 slomerulosclerosis 8
1POA1	Ipolipoprotein A-1Vichols Genomics 8:318, 1990ID	Aultiple + 107680 classifications
1RHGAP24	ChoGTPaseactivatinglkilesh J Clin Invest 121:4127,1Dvrotein 242011	VA <sup>°</sup> 610586
NF2	nverted formin, FH2 and Brown Nat Genet 42:72, 2010 1D VH2 domain containing	Flomerulosclerosis, ±613237 ocal segmental, 5
MX1B	IMHomeobox)reyer Nat Genet 19:47 19981DFranscription Factor 1 Beta	Vail-patella ± 161200 yndrome
<i>AEFV</i>	<i>AEFV</i> Innate Immunity?ergman Am J Med 45:601,1D <i>Cegulator, Pyrin</i> 968	Familial ±134610 Aediterranean fever, 1D

ЛҮН9	Ayoson heavy chain conmuscle	9,1eath AJHG 69:1033, 2001 1D	<i>Aacrothrombocytope</i> ± 155100 via and granulocyte nclusions with or vithout nephritis or ensorineural vearing loss
PODXL	Podocalyxin	3arua Kidney Int 85:124, 2014 1D	VA \$602632
SLC37A4	Solute Carrier Family Glocose-6-Phosphate Transporter), Member 6	37Nordlie J Biol Chem 258:9739, '983	Flycogen storage <sup>‡</sup> 232240 lisease Ic
TRPC6	Transient receptor potents ation channel, subfamily nember 6	ialVinn Science 308:1801, 2005 1D C,	Flomerulosclerosis, ±603965 Tocal segmental, 2
VT1	Vilms Tumor 1	<i>Aelo J Clin Endocrinol MetablD</i> 37:2500, 2002	Frasier syndrome ±136680
KBKAP	nhibitor of kappa lig volypeptide gene enhancer 3 cells, kinase compl vssociated protein	ghtInderson AJHG 68:753, 2001 IR/AD in Jex	Familial ± 223900 lysautonomia
√XF5	Vuclear RNA export factor	5 Esposito Hum Mol Genet(L ?2:3654, 2013	VA \$300319
1POE	lpolipoprotein E	Dikawa JASN 8:820, 1997 (L	<i>Aultiple '107741</i> <i>lassifications</i>
IPOL1	lpolipoprotein L-1	Parsa NEJM 369:2183, 2013 Jnknown	Susceptibility to <sup>4</sup> 612551 vondiabetic rnd-stage renal lisease, Susceptibility to focal egmental slomerulosclerosis 4
GPC5	Hypican 5	Dkamoto Nat Genet 43:459, Jnknown 2011	VA \$602446
SYNPO	lynaptopodin	Bierzynska Kidney Int 91:937, Jnknown 2017	<i>√A</i> °608155
.PIN1	ipin1	Christensen Danish Med. Bull.1R 20: 112-115, 1983	<i>Ayoglobinuria, acute ecurrent, autosomal ecessive; renal `ailure</i>
CNJ5	Potassium channel, inqarc ectifying, subfamily namber 5	llyAulatero Hypertension 59:1D J,'35-240, 2012	Typeraldosteronism, 500734 amilial, type III
1MN	1mnion-associated ransmembrane protein	De Filipo Ital. J. Pediat. 39: 58,1R 2013	merslund-Grasbeck 105799 yndrome 2

1POC2	lpolipoprotein C-II	Cashyap Atherosclerosis, 35(1),1R '9–40, 1980	Vephrotic syndrome 608083
<i>G6PC1</i>	<i>Flucose-6-Phosphatase</i> <i>Catalytic Subunit 1</i>	Carthi Gene, 700, 7–16, 2019 IR	Tlycogen storage√A lisease Ia
COQ8B	Coenzyme Q8B	1shraf J. Clin. Invest. 123:1R 5179-5189, 2013	Vephrotic syndrome,15567 vpe 9
1LG13	1LG13 JDP-N-Acetylglucosaminyl ansferase subunit	Esposito Human molecular/LR It jenetics, 22(18), 3654–3666, 2013	Focal segmental <sup>1</sup> 00776 slomerulosclerosis
76	Complement component 6	eddy J. Clin. Invest. 53:1R 44-553, 1974	C6 deficiency 217050
ELP1	Slongator complex protein	1 Inderson Am J Hum Genet.1D/AR i8(3):753–758, 2001	Dysautonomia, 103722 'amilial

*AR*, autosomal recessive; *AD*, autosomal dominant; Unknown, mode of inheritance not clearly characterized; *NA*, not available; *OMIM*, Online Mendelian Inheritance in Man; *XL*; *X*-linked; #, phenotype MIM number; \* gene/locus MIM number if not phenotype MIM number available; +, gene and phenotype combined.

Supplementary	Table S3: genes	that represen	t monogenic d	causes of human	isolated CA	KUT. if mutated
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Jene	<sup>•</sup> rotein	leference	10de nheritance	ofhenotype	OMIM #
1CE	Ingiotensin I-converting enzyme	<i>Fribouval Nat (</i> 17:964, 2005	GenetlR	Renal lysgenesis	tubular <sup>‡</sup> 267430
lGT	lngiotensinogen	<i>Fribouval Nat (</i> 7:964, 2005	GenetlR	Renal lysgenesis	tubular <sup>‡</sup> 267430
IGTR1	Ingiotensin II receptor, type 1	Fribouval Nat ( 7:964, 2005	GenetlR	Renal lysgenesis	tubular <sup>‡</sup> 267430
CHRM3	<i>Auscarinic acetylcholine recepto</i> <i>13</i>	nrVeber AJHG 19 2011	:634,1R	Prune yndrome	belly <sup>‡</sup> 100100
ETV4	ETS translocation variant 4, EL nhancer binding protein	A <sup>°</sup> hen IJPCH 4:61, 2	2016 IR	√A	<i>600711</i>
FRAS1	Extracellular matrix protei FRASI	nCohl JASN 25: 2014	1917,1R	Fraser synd	drome 1±219000
FREM1	7RASI related extracellular matri vrotein 1	ixohl JASN 25: '014	1917,1R	∫anitoba >culotricho yndrome	±248450 anal
FREM2	<sup>7</sup> RAS1 related extracellular matri protein 2	ixCohl JASN 25: 2014	1917,1R	Fraser sync	lrome 2‡617666
GRIP1	<i>Flutamate receptor interactin</i> vrotein l	gʻohl JASN 25: 2014	1917,1R	Fraser sync	lrome 3±617667
IPSE2	<i>Heparanase 2 (Inactive)</i>	<i>Bulum Nephron 13</i> 2015	0:54,1R	Jrofacial yndrome I	ŧ236730

TGA8	ntegrin a8	Humbert AJHG1R 89:1260, 2014	Renal ±191830 vypodysplasia/ vplasia 1
₹EN	Renin	Fribouval Nat GenetlR 7:964, 2005	Renal tubular <sup>‡</sup> 267430 lysgenesis
"RAP1	<i>Heat-shock protein 75 (also know</i> s TNF receptor-associate wotein 1)	vn`aisawat Kid Int 85:880,1R ed`014	VA <sup>°</sup> 606219
7GF20	Fibroblast Growth Factor 20	3arak Dev Cell 22:1191,1R 2012	Renal ±615721 ypodysplasia/ yplasia 2
3MP4	<i>Sone morphogenic protein 4</i>	Veber JASN 19:891,1D 2008	Aicrophthalmia, ↓607932 yndromic 6
CHD1L	Chromodomain helicase DN vinding protein 1-like	<i>IA</i> 3rockschmidt NDT\D ?7:2355, 2012	VA <sup>6</sup> 13039
CRKL	CRK Like Proto-Oncogen daptor protein	ne, .opez-Rivera NEJMID 176: 742, 2017	VA * 602007
)STYK	<i>Jual serine/threonine and tyrosi</i> <i>vrotein kinase</i>	ne'anna-Cherchi NEJMID 869:621, 2013	Congenital ±610805 momalies of idney and urinary ract 1
EYA I	Eyes absent homolog 1	1bdelhak Nat Genet1D 5:157, 1997	Branchiootorenal ±113650 yndrome 1, with wr without vataracts
7ATA3	<i>GATA binding protein 3</i>	Pandolfi Nat GenetlD 1:40, 1995; Van Esch Vature 406:419, 2000	Hypoparathyroidis‡146255 n, sensorineural leafness, and enal dysplasia
GREBIL	Growth Regulation By Estrogen Breast Cancer 1 Like	Introphy Genetics1D 207:215, 2017, Vanna-Cherchi AJHG 201:1034, 2017	Renal ±617805 vypodysplasia/ vplasia 3
INF1B	INF homeobox B	indner Hum Mol GenetlD 24:263, 1999	Renal cysts and <sup>t</sup> 137920 liabetes syndrome
AUC1	Aucin 1	Cirby Nat Genet 45:299,1D 2013	Лedullary cystic <sup>t</sup> 174000 idney disease 1
<i>√FIA</i>	vuclear Factor 1/A	<i>αο Eur J Med Genet</i> Ω 7:65, 2014	Brain £613735 nalformations vith or without vrinary tract lefects

VRIP1	<i>Juclear Receptor Interactiv</i> Protein I	agʻivante JASN 28:2364,1D '107	VA '602490
?AX2	Paired box 2	Sanyanusin Hum MollD Fenet 4:2183, 1995	Papillorenal ±120330 yndrome
<i>₽BX1</i>	PBX Homeobox 1	Heidet JASN 28:2901,1D '017	Congenital ±617641 nomalies of idney and urinary ract syndrome vith or without hearing loss, hbnormal ears, or levelopmental lelay
`PP3CA	Protein Phosphatase 3, Catalyt Subunit, Alpha Isoform	ic⁄lizuguchi Hum MollD Genet 27:1421, 2018	Irthrogryosis, left palate, raniosynostosis, und impaired ntellectual levelopment
₹ET	Proto-oncogene tyrosine-prote inase receptor Ret	inškinner AJHG 82:344,1D 2008	Aultiple OMIM <sup>®</sup> 164761 classifications
ROBO2	<i>Roundabout, axon guidan</i> <i>eceptor, homolog 2 (Drosophila)</i>	ceIwang Hum GenetlD '34:905, 2015; Lu IJHG 80:616, 2007	<sup>7</sup> esicoureteral <sup>€</sup> 610878 eflux 2
ALLI	Sal-like protein 1 (also known a palt-like transcription factor 1)	asCohlhase Nat GenetlD '8:81, 1998	Fownes-Brocks ±107480 yndrome 1
SIX1	IX homeobox 1	Ruf PNAS 101: 8090,1D 2004	<i>Branchio-otic £608389</i> <i>yndrome 3</i>
VIX2	IX homeobox 2	Veber JASN 19:891,1D '008	VA <sup>604994</sup>
SIX5	NX homeobox 5	Hoskins AJHG 80:800,1D 2007	3ranchiootorenal ±610896 yndrome 2
SLIT2	ilit homolog 2	Hwang Hum GenetlD '34:905, 2015	VA '603746
OX17	ranscription factor SIX-17	<i>Fimelli Hum Mut\D</i> 11:1352, 2010	∕esicoureteral ±613674 eflux 3
RGAP1	<i>LIT-ROBO Rho GTPa.</i> <i>activating protein 1</i>	seIwang Hum GenetlD '34:905, 2015	VA \$606523
"BX18	-Box transcription factor	'ivante AJHG 97:291,1D '015	Congenital ±143400 momalies of idney and urinary ract 2

NXB	<sup>r</sup> enascin XB	Fibadegesin     JASNID       24:1313, 2013	′esicoureteral ⊧615963 eflux 8
JPK3A	Jroplakin 3A	'enkins JASN 16:2141,1D '005	VA <sup>°</sup> 611559
VNT4	<sup>2</sup> rotein Wnt-4	<i>Siason-Lauber NEJMID</i> <i>Siason-Lauber NEJMID</i> <i>Siaso</i>	Aullerian aplasia‡158330 ınd ıyperandrogenism
CAL1	Inosmin I	Hardelin PNAS 89:8190,(L '992	<i>Hypogonadotropic</i> <sup>±</sup> 308700 <i>ypogonadism</i> 1 <i>vith or without</i> <i>nosmia</i> <i>Kallmann</i> <i>yndrome</i> 1)
FAM58A	Family with Sequence Similarit 8 Member A	yJnger Nat Genet(LD 40:287, 2008	Foe Syndactyly,± 300707 Felecanthus, and Ingiogenital and Renal Malformations

Supplementary Table S4: genes that represent monogenic causes of human syndromic CAKUT, if mutated.

Jene	rotein	Reference	/lode nheritanc	of henotype e	)MIM umber
33GALTL	Beta 3-Glucosyltransferase	Lesnik Oberstein A '9:562, 2006	JHGAR	'eters-plus yndrome	<sup>!</sup> 261540
3SCL2	3SCL2, Seipin Lipid 3iogenesis Associated	DropletIaghighi Clin Gene 34, 2016	t 89:AR	Aultiple lassifications	<sup>£</sup> 608594 <sup>£</sup> 269700 <sup>£</sup> 600794 <sup>£</sup> 615924 <sup>£</sup> 270685
CD151	CD151 Molecule(RaphGroup)	BloodCaramatic <i>H</i> 04:2217, 2004	3loodAR	JA	602243
CD96	D96 Molecule	Kaname <i>AJHG</i> 81 2007	:835,AR	2 syndrome	<sup>!</sup> 211750
CHRNG	Cholinergic Receptor M Gamma Subunit	Nicotinic/ogt J Med Genet 4 2012	9:21,AR	Escobar syndrom	ne <sup>!</sup> 265000
CISD2	CDGSH Iron Sulfur Domain	1 2 Amr <i>AJHG</i> 81:673, 2	007 AR	Volfram yndrome 2	<sup>!</sup> 604928

CTU2	Cytosolic Thiouridylase, subunit 2	Shaheen <i>AJMG</i> 170:3222,\R :016	Aicrocephaly, <sup>1</sup> 618142 acial lysmorphism, enal agenesis, and mbiguous genitalia syndrome
<u>77721</u>	Cytochrome P450 Family 21	Martul Arch Dis ChildAR 55:324, 1980	Iyperandrogenis <sup>1</sup> 201910 n, nonclassic ype, due to 1-hydroxylase leficiency
)ACH1	Dachshund Family Transcriptic	michild <i>NDT</i> 28:227, 2013 AR	VA 603803
OHCR7	'-Dehydrocholesterol Reductase	.öffler <i>AJHG</i> 13;95:174,\R :000	Smith-Lemli-Opit <sup>§</sup> 270400 Syndrome
DIS3L2	DIS3 Like 3'-5' Exoribonuclease 2	Astuti <i>Nat Genet</i> AR ;;44:277, 2012	erlman syndrome <sup>1</sup> 267000
EMG1	MG1, N1-Specific Pseudouridir Aethyltransferase	heArmistead AJHG 84:728,AR 2009	Bowen-Conradi <sup>£</sup> 211180 yndrome
ERCC8	Excision repa ross-complementing, group 8	ir3ertola J Hum GenetAR 11:701, 2006	Cockayne <sup>±</sup> 216400 yndrome, type A
ESCO2	Establishment Of Sister Chromatic Cohesion N-Acetyltransferase 2	id/ega <i>J Med Genet</i> 47:30,\R :010	Roberts syndrome <sup>1</sup> 268300
ETFA	Electron Transfer Flavoprotei	in.ehnert <i>Eur J Pediatr</i> \R 39:56, 1982	ilutaric acidemia <sup>!</sup> 231680 IA
ETFB	Electron Transfer Flavoprotein Ber Subunit	ta.ehnert <i>Eur J Pediatr</i> AR 39:56, 1982	ilutaric acidemia <sup>1</sup> 231680 IB
ETFDH	Electron Transfer Flavoprote	in.ehnert <i>Eur J Pediatr</i> \R 39:56, 1982	ilutaric acidemia <sup>!</sup> 231680 IC
<sup>7</sup> ANCA	Panconi Anemia Complementatio Broup A	noenje & Patel Nat RevAR Genet 2:466, 2001	Fanconi anemia, <sup>1</sup> 227650 omplementation group A
FANCB	Fanconi Anemia Complementatio Froup B	n/acCauley <i>AJMG</i> \R 55A:2370, 2011	Fanconi anemia, <sup>1</sup> 300514 complementation group B
FANCD2	Fanconi Anemia Complementatio	nKalb <i>AJHG</i> 80:895, 2007 AR	anconi anemia, <sup>1</sup> 227646 complementation group D2
FANCE	Fanconi Anemia Complementatio Froup E	onVegner <i>Clin Genet</i> AR 50:479, 1996	Fanconi anemia, <sup>1</sup> 600901 complementation group E

<sup>7</sup> ANCI	Panconi Anemia Complementation Broup I	on¦avage <i>AJMG</i> 170A:386,\R 2015	Fanconi anemia, <sup>£</sup> 609053 complementation group I
FANCL	Panconi Anemia Complementation Broup L	on/etro <i>Hum Mutat</i> 36:562, AR 2015	Fanconi anemia, <sup>1</sup> 614083 complementation group L
FAT4	AT Atypical Cadherin 4	Alders     Hum     GenetAR       33:1161, 2014	/an Maldergem <sup>!</sup> 615546 yndrome 2
7OXP1	Forkhead Box P1	Bekheirnia <i>Genet Med</i> AR 9:412, 2017	NA 605515
HES7	Ies Family BHLH Transcriptio	onsparrow <i>Hum Mol Genet</i> AR 7:3761, 2008	JA 608059
HYLS1	IYLS1, Centriolar And Ciliogenes Associated	sis'aetau <i>J Neuropathol Exp</i> \R Veurol 67:750, 2008	Iydrolethalus <sup>1</sup> 236680 yndrome
СК	ntertinal cell kinase	ahiry <i>AJHG</i> 84:822,AR	JA 612325
FT46	ntraflagellar Transport 46	Lee Dev Biol 400:248,AR 2015	Short-rib thoracic <sup>£</sup> 617102 lysplasia 16 with or without olydactyly
FT74	ntraflagellar Transport 74	Cevik <i>PLoS GeneT</i> AR P:e1003977, 2013	Bardet-Biedl5617119yndrome 20
TGA3	ntegrin Subunit Alpha 3	(alcin <i>Hum Mol Genet</i> AR (4:3679, 2015	nterstitial lung <sup>4</sup> 614748 lisease, nephrotic yndrome, and pidermolysis vullosa, congenital
'AM3	unctional Adhesion Molecule 3	Aochida <i>AJHG</i> AR 0;87:882, 2010	Hemorrhagic <sup>1</sup> 613730 lestruction of the vrain, ubependymal alcification, and ataracts
MNA	.amin A/C	Clupa <i>Endocrine</i> 36:518,AR 2009	Aultiple OMIM 150330 lassification
.RIG2	Leucione rich repeats an mmunoglobulin like lomains containing protein 2	ndituart <i>AJHG</i> 92:259,AR :013	Jrofacial yndrome 2 ± 615112
RP2	.DL Receptor Related Protein 2	Cantarci <i>Nat Genet</i> AR 9:957, 2007	Jonnai-Barrow <sup>£</sup> 222448 yndrome
.RP4	.DL Receptor Related Protein 4	.i Am <i>J Hum Genet</i> AR 66:696, 2010	Cenani-Lenz <sup>£</sup> 212780 yndactyly yndrome

AESP2	Лesoderm Posterior BHL Transcription Factor 2	HJeorge-Abraham <i>AJMG</i> \R \ 158A:1971, 2012	JA 605195
AKS3	Aeckel Syndrome Type 3 Protein	3aala <i>AJHG</i> 80:186, 2007 \R	Aeckel syndrome <sup>t</sup> 607361
PEX5	eroxisomal Biogenesis Factor 5	Sundaram Nat Clin PractAR Fastroenterol Hepatol E:456, 2008	Peroxisome ±214110 viogenesis lisorder 2A Zellweger)
·MM2	hosphomannomutase 2	Iorslen <i>Arch Dis Child</i> AR 6:1027, 1991	Congenital <sup>§</sup> 212065 lisorder of glycosylation, type a
POCIA	OC1 centriolar protein	Shaheen <i>AJHG</i> 91:330,AR 2012	Short stature, <sup>£</sup> 614813 inychodysplasia, acial lysmorphism, and iypotrichosis
'RODH	Proline dehydrogenase 1	Perry Ann Hum GenetAR 1:401, 1968	Iyperprolinemia, ±239500 Type 1
PROK2	Prokineticin 2	Aadan <i>Mol Genet Metab</i> \R <i>≷ep</i> 12:57, 2017	Iypogonadotropic <sup>£</sup> 610628 ypogonadism 4 vith or without nosmia
₹ <i>AD51C</i>	CAD51 Paralog C	/az Nat Genet 42:406,AR 2010	Fanconi Anemia, <sup>1</sup> 613390 Complementation Froup O
RECQL4	RecQ Like Helicase 4	Siitonen <i>Eur J Hum Genet</i> AR 7:151, 2009	3aller-Gerold <sup>1</sup> 218600 yndrome
₹MND1	Required for Meiotic Nucle Division 1 Homolog	ar`aylor <i>JAMA</i> 312:68,AR 2014	Combined ± 614922 Dxidative Phosphorylation Deficiency 11
COR2	Receptor Tyrosine Kinase Lil Drphan Receptor 2	keViens <i>Clin Genet</i> 37:481,AR 990	Kobinow½268310yndrome
RPS19	Ribosomal Protein S19	Ioefele <i>Pediatr Nephrol</i> AR 5:1255, 2010	JA 603474
CARF2	Scavenger Receptor Class Member 2	FAnastasio <i>AJHG</i> 87:553,AR 2010	/an den <sup>!</sup> 600920 Ende-Gupta yndrome
TRA6	Stimulated By Retinoic Acid 6	Golzio <i>AJHG</i> 80:1179,AR 2007	Aicrophthalmia, ±601186 yndromic 9

			Aicrophthalmia, solated, with oloboma 8
"MCO1	ransmembrane And Coiled-Coi Domains 1	Kin <i>PNAS</i> 107:258, 2010 AR	Craniofacial <sup>1</sup> 213980 lysmorphism, keletal anomalies, nd mental etardation yndrome
JBR1	Jbiquitin Protein Ligase E3 Component N-Recognin 1	3/anlieferinghen GenetAR Couns 14:105, 2003	ohanson-Blizzard <sup>1</sup> 243800 yndrome
°EXI	Peroxisomal Biogenesis Factor 1	Crane <i>Hum Mutat</i> 26:167,AR 2005	Peroxisome ±214100 piogenesis lisorder 1A Zellweger)
PIGL	hosphatidylinositol Glycan Anchor Biosynthesis Class L	ršchnur <i>AJMG</i> 72:24,AR 997	CHIME syndrome <sup>1</sup> 280000
чGO	Phosphatidylinositol Glycan Anchor Biosynthesis Class O	Krawitz <i>AJHG</i> 91:146,AR 2012	Iyperphosphatasi ±614749 with mental etardation yndrome 2
PIGN	'hosphatidylinositol Glycan Anchor Biosynthesis Class N	r)hba NeurogeneticsAR 5:85, 2014	Aultiple± 614080ongenitalnomalies-hypotoiia-seizuresyndrome 1
PIGT	<sup>v</sup> hosphatidylinositol Glycan Anchor Biosynthesis Class T	Nakashima <i>Neurogenetics</i> AR 5:193, 2014	Multiple* 615398ongenitalnomalies-hypotoiia-seizuresyndrome 3
2IGV	Phosphatidylinositol Glycan Anchor Biosynthesis Class V	Horn <i>Eur J Hum Genet</i> AR 2:762, 2014	VA 610274
PIGY	Phosphatidylinositol Glycan Anchor Biosynthesis Class Y	rlkovski <i>Hum Mol Genet</i> AR '4:6146, 2015	Iyperphosphatasi <sup>1</sup> 616809 with mental etardation yndrome 6
PTF1A	<sup>2</sup> ancreas Specific Transcriptior <sup>2</sup> actor, 1a	Burung <i>Mol Med Rep</i> \R 2:1579, 2015	JA <sup>607194</sup>
SLX4	SLX4 Structure-Specific Endonuclease Subunit	Stoepker <i>Nat Genet</i> AR -3:138, 2011	<sup>2</sup> anconi anemia, <sup>1</sup> 613951 complementation group P
"XNL4A	Thioredoxin-Like 4A	Hing Am J Med Genet AAR       40:804, 2006	Burn-McKeowen <sup>§</sup> 608572 Syndrome

VFS1	Volframin ER Transmembran Jlycoprotein	esalih <i>Acta Paediatr Scand</i> AR 50:567, 1991	Volfram <sup>1</sup> 222300 yndrome 1
VNT3	Vnt Family Member 3	Viemann <i>AJHG</i> 74:558,AR 2004	Petra-amelia <sup>1</sup> 273395 yndrome 1
RCC4	ζ-Ray Repair Cross Complementing	gЛurray <i>Am J Hum Genet</i> AR )6:412, 2015	Short Stature, <sup>£</sup> 616541 Aicrocephaly, And Endocrine Dysfunction
ZMPSTE24	Zinc Metallopeptidase STE24	Zhen     AJMG     AAR       49A:1550, 2009	Restrictive <sup>1</sup> 275210 lermopathy, lethal
1CTB	Actin Beta	tivière <i>Nat Genet</i> 44:440, AD 2012	3araitser-Winter ±243310 yndrome 1
1CTG1	Actin Gamma 1	livière Nat Genet 44:440, AD 2012	Baraitser-Winter ±243310 yndrome 1
1 <i>IFM3</i>	Apoptosis Inducing Factor Aitochondria Associated 3	r, opez-Rivera <i>NEJM</i> AD 76:742, 2017	JA 617298
1 <i>RID1B</i>	\T-Rich Interaction Domain 1B	evy J Med Genet 28,AD 991	Coffin-Siris! 135900yndrome 1
1TXN10	Ataxin 10	Aatsuura     Nat     GenetAD       :6:191, 2000	Spinocerebellar ±603516 taxia 10
31CC1	3icC Family RNA Binding Protein	n\'raus <i>Hum Mutat</i> 33:86,\D 2012	Renalcystic/! 601331lysplasia
3MP7	Bone Morphogenetic Protein 7	Iwang     Kidney     IntAD       55:1429, 2014	JA 112267
3RAF	3-Raf Proto-Oncogene Gerine/Threonine Kinase	e, arkozy <i>Hum Mutat</i> AD 60:695, 2009	Cardiofaciocutane ± 115150 vus syndrome
CDC5L	Cell Division Cycle 5 Like	Groenen GenomicsAD 49:218, 1998	JA 602868
CREBBP	CREB Binding Protein	Canjilal <i>J Med Genet</i> AD 19:669, 1992	Rubinstein-Taybi ±180849 yndrome 1
)ACT1	Dishevelled Binding Antagonist O Beta Catenin 1	fVebb <i>Hum Mutat</i> 38:373,AD :017	Cownes-Brocks ± 617466 yndrome 2
EP300	E1A Binding Protein P300	toelfsema <i>AJHG</i> 76:572, AD 2005	Rubinstein-Taybi½ 613684yndrome 2
ESRRG	Estrogen Related Receptor Gamma	Harewood     PLoS     OneAD       ::e12375, 2010	VA 602969
7BN1	ibrillin 1	JokhmafshanPediatrADVephrol 32:565, 2017	Marfan syndrome ±154700

₹GFR1	ibroblast growth factor receptor 1	<sup>°</sup> arrow <i>AJHG</i> 140A:537,4D 2006	Aultiple     OMIM!     615465       lassifications     !     147950       !     123150     !       !     166250     !       !     101600     !       !     190440     !
GFR3	ibroblast growth factor receptor 3	Rohmann <i>Nat Genet</i> AD 8:495, 2006	ADD syndrome <sup>1</sup> 149730
7GF10	ibroblast Growth Factor 10	<i>A</i> ilunsky <i>Clin Genet</i> AD 9:349, 2006; Bamforth 1 <i>JMG</i> 43:932, 1992	ADD syndrome <sup>1</sup> 149730
7GF8	ibroblast Growth Factor 8	Falardeau <i>J Clin Invest</i> AD 18:2822 2008	Iypogonadotropic <sup>1</sup> 612702 Iypogonadism 6 vith or without nosmia
FGFR2	ibroblast Growth Factor Receptor	2.eHeup <i>Eur J Pediatr</i> AD 54:130, 1995	Aultiple OMIM 176943 lassifications
GFRL2	Forkhead Box C1	eHeup <i>Eur J Pediatr</i> AD 54:130, 1995	Antley-Bixler <sup>1</sup> 207410 yndrome without genital anomalies yr disordered teroidogenesis
FMN1	°ormin 1	Dimitrov     J     Med     GenetAD       :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010     :7:569, 2010	JA 136535
7OXF1	Forkhead Box F1	Hilger <i>Hum Mutat</i> AD 6:1150, 2015	Alveolar capillary <sup>£</sup> 265380 lysplasia with nisalignment of ulmonary veins
GDF3	Growth Differentiation Factor 3	Karaca     AJMG     A\D       67A:2795, 2015     674	Klippel-Feil± 613702yndrome 3
GDNF	ilial cell line derived neurotrophi	c'ini Prato <i>Medicine</i> AD <i>Baltimore)</i> 88:83, 2009	Susceptibility <sup>1</sup> 613711 Iirschsprung Disease
GFRA1	DNF Family Receptor Alpha 1	ChatterjeeHumGenetAD31:1725, 2013	JA 601496
<i><b>FLI2</b></i>	HI Family Zinc Finger 2	Carmichael <i>J Uro</i> AD 90:1884, 2013	Culler-Jones <sup>£</sup> 615849 yndrome, <sup>£</sup> 610829 Holoprosencephal
HOXA13	Iomeobox A13	Ialal AJMG 30:793, 1998\D	Iand-foot-uterus <sup>1</sup> 140000 yndrome
HOXD13	Iomeobox D13	Jarcia-Barceló     AJMGAD       1146A:3181, 2008	JA 142989

'AG1	agged 1	Kamath Nat Rev NephroAD 9:409, 2013	Alagille syndrome <sup>t</sup> 118450
CAT6B	.ysine Acetyltransferase 6B	Campeau <i>AJMG</i> 90:282, AD 2012	Genitopatellar <sup>£</sup> 606170 yndrome
CTD1	Potassium Channel Tetramerizatio Domain Containing 1	nAarneros AJMG 92:621,AD 2013	Scalp-ear-nipple <sup>1</sup> 181270 yndrome
CNH2	Potassium Voltage-Gated Channe Subfamily H Member 2	elčaselli <i>AJMG</i> 146A:1195,AD 2008	scalp-ear-nipple 152427 yndrome
CRAS	CRAS Proto-Oncogene, GTPase	Schubbert <i>Nat Gene</i> \D 8:331, 2006	Joonan syndrome <sup>1</sup> 609942
MX1B	JM Homeobox Transcriptio	nDreyer <i>Nat Genet</i> 19:47,AD 998	Vail-patella <sup>1</sup> 161200 yndrome
.PP	IM Domain Containing Preferre Translocation Partner In Lipoma	dIernández-García <i>AJMG</i> AD 1158A:1785, 2012	JA 600700
AP2K1	<i>A</i> itogen-activated protein kinas	eschulz <i>Clin Genet</i> 73:62,AD 2007	Cardiofaciocutane <sup>£</sup> 615279 ous syndrome 3
AP2K2	Aitogen-activated protein kinas	eschulz <i>Clin Genet</i> 73:62,AD 2007	Cardiofaciocutane <sup>£</sup> 615280 ous syndrome 4
1LL2/ XMT2D	Ayeloid/LymphoidCAixed-Lineage Leukemia Protein 2	r3anka <i>Eur J Hum Genet</i> AD :0:381, 2012	Cabuki syndrome! 147920
ANX1	Aotor Neuron and Pancrea Iomeobox 1	us\shcraft J Pediatr Surg\D 1:691, 1974	Currarino <sup>!</sup> 176450 Syndrome
AYCN	AYCN     Protooncogene,     bHLl       Transcription     Factor	H <i>M</i> arcelis <i>Hum Mut</i> AD 9:1125, 2006	reingold ±164280 yndrome 1
VOTCH2	Jotch 2	Kamath Nat Rev NephroAD 9:409, 2013	Alagille syndrome <sup>!</sup> 610205 !, Hajdu-Cheney! 102500 yndrome
'KD1	Polycystin 1, Transient Receptor Potential Channel Interacting	orcossetti JASN 18:2143, AD 2007	'olycystic kidney <sup>!</sup> 173900 lisease 1
°KD2	Polycystin 2, Transient Receptor Potential Cation Channel	orcossetti JASN 18:2143, AD 2007	olycystic kidney <sup>!</sup> 613095 lisease 2
PROKR2	Prokineticin Receptor 2	Sarfati Front Horm Res\D 9:121, 2010	Iypogonadotropic <sup>1</sup> 244200 Iypogonadism 3 vith or without nosmia
PTPN11	Protein Tyrosine Phosphatase √on-Receptor Type 11	e,3ertola <i>AJMG</i> 130A:378,4D 2004	LEOPARD <sup>1</sup> 151100 yndrome 1
₹AF1	Caf-1Proto-OncogeneSerine/Threonine Kinase	e, azzaque <i>Nat Genet</i> AD 9:1013, 2007	Joonan syndrome <sup>1</sup> 611553
₹AII	Retinoic Acid Induced 1	/ilboux     PLoS     One\D       b:e22861, 2011     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0<	Smith-Magenis <sup>1</sup> 182290 yndrome

RPL26	tibosomal protein L26	Gazda <i>Hum Mutat</i> AD 3:1037, 2012	Diamond-Blackfa <sup>±</sup> 614900 1 anemia 11
₹PS26	tibosomal protein S26	Gripp Am J Med Genet AAD 64A:2240, 2014	Diamond-Blackfa <sup>£</sup> 613309 1 anemia 10
SALL4	Spalt Like Transcription Factor 4	CohlhaseADGeneReviews®BookSection, 1993	Duane-radial ray <sup>±</sup> 607323 yndrome
EMA3A	Semaphorin 3A	⟨oung Hum Reprod\D 27:1460, 2012	Iypogonadotropic <sup>1</sup> 614897 Iypogonadism 16 vith or without nosmia
SEMA3E	emaphorin 3E	Lalani <i>J Med Genet</i> AD 1:e94, 2004	CHARGE <sup>1</sup> 214800 yndrome
ETBP1	ET Binding Protein 1	Schinzel <i>AJMG</i> 1:361, AD 978	Schinzel-Giedion ±269150 nidface retraction yndrome
SHH	Sonic Hedgehog	urie AJMG 35:286, 1990\D	Ioloprosencephal ± 142945
<i>F3B4</i>	Splicing Factor 3b Subunit 4	Bernier <i>AJMG</i> 90:925,AD 2012	Acrofacial½ 154400lysostosis1,Jager type
NAP29	Synaptosome Associated Protein 2	9 .opez-Rivera <i>NEJM</i> AD 76:742, 2017	Di George 604202 yndrome
SOS1	OS Ras/Rac Guanine Nucleotic Exchange Factor 1	de'errero Eur J Med GenetAD 11:566, 2008	Joonan syndrome <sup>1</sup> 610733
OX9	SRY-Box 9	Airik     Hum     Mol     Genet\D       9:4918, 2010     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$     \$ </td <td>Campomelic <sup>£</sup> 114290 lysplasia</td>	Campomelic <sup>£</sup> 114290 lysplasia
RCAP	Snf2 Related CREBBP Activate Protein	orłood <i>AJHG</i> 90:308, 2012\D	iloating-Harbor 136140 yndrome
"BX1	-Box 1	Sujat AJMG A 140:1601,AD 2006	Di George <sup>t</sup> 188400 yndrome
"BX3	2-Box 3	Aeneghini Eur J MedAD Genet 49:151, 2006	Лnar-mammary <sup>±</sup> 181450 yndrome
"FAP2A	Transcription Factor AP-2 Alpha	Ailunsky AJHG 82:1171,AD 2008	3ranchiooculofaci <sup>1</sup> 113620 1 syndrome
<sup></sup> P63	Cumor Protein P63	Celli Cell 99:143, 1999 AD	Aultiple OMIM 603273 lassifications
TRPS1	Zinc finger transcription factor Trichorhinophalangeal syndrome	r; asic <i>Ren Fail</i> 36:619,AD 2014	richorhinophalan <sup>؛</sup> 190350 eal syndrome ؛ 190351
"SC1	uberous Sclerosis 1	Curatolo Lancet 372:657,AD 2008	Cuberous! 191100clerosis-1

"SC2	Suberous Sclerosis 2	Kumar <i>Hum Mol Genet</i> AD 1:1471, 1995	luberous ± 613254 clerosis-2
TWIST2	Wist Family BHLH Transcription	nitevens <i>AJMG</i> 107:30, \D :002	Ablepharon-macro <sup>1</sup> 200110 tomia syndrome
VNT5A	Vnt Family Member 5A	Roifman     Clin     GenetAD       57:34, 2015;     Person     Dev       Dyn 239:327, 2010     Dev	tobinow! 180700yndrome
GDF6	Browth Differentiation Factor 6	Cassabehji <i>Hum Mutat</i> AD/ AR 29:1017, 2008	Aultiple OMIM 601147 lassifications
<i>7LI3</i>	3LI Family Zinc Finger 3	Cain <i>PLoS One</i> 4:e7313, \D/ AR 2009	Aultiple OMIM 165240 lassifications
°CSK5	roprotein Convertase Subtilisin a čexin Type 5	&Jakamura <i>BMC Res</i> \D/ AR <i>Votes</i> 8:228, 2015	JA 600488
PTEN	hosphatase And Tensin Homolog	Reardon J Med GenetAD/ AR8:820, 2001	Aultiple OMIM 601728 lassifications
<i>PS24</i>	Ribosomal Protein S24	Zetgin     Turk     J     Pediatr\D/ AR       6:239, 1994     6:239, 1994     6:239, 1994     6:239, 1994	Nase-Smith602412yndrome
VANGL1	/ANGL Planar Cell Polarity Protei	n3artsch <i>Mol Syndromol</i> AD/ AR ::76, 2012	Caudal regression <sup>t</sup> 600145 yndrome
IXINI	Axin 1	Dates AJHG 79:155, 2006)e novo	Caudal duplication <sup>£</sup> 607864 nomaly
<del>1</del> 19	I19,ImprintedMaternallExpressedTranscript(Non-ProteinCoding)ImprintedImprinted	lyłur <i>PNAS</i> 113:10938, <i>)e novo</i> n:016	Beckwith-Wiedem <sup>1</sup> 130650 nn syndrome
CNQ10T1	CNQ1 Opposite Strand of Antisense Transcript 1 (Non-Protein Coding)	& Thiesa Hum Mol Genet De novo in 1:10, 2012	Beckwith-Wiedem <sup>1</sup> 130650 nn syndrome
VIPBL	VIPBL, Cohesin Loading Factor	Rohatgi     AJMGDe novo       52A:1641, 2010	Cornelia de Lange <sup>!</sup> 122470 yndrome 1
CDKNIC	Cyclin Dependent Kinase Inhibito	orAussa <i>Pediatr NephroDe novo</i> .7:397, 2012	Beckwith-Wiedem <sup>1</sup> 130650 nn syndrome
CHD7	Chromodomain Helicase DNA Binding Protein 7	A anssenHumMutat)e novo3:11492012	CHARGE ±214800 yndrome
IMER1	APC Membrane Recruitmen Protein 1	nt'ellegrino AJMG 16:159,(L 997	Osteopathia striata <sup>1</sup> 300373 vith cranial clerosis
ITP7A	ATPase Copper Transporting Alpha	1 Julpe Nat Genet 3:7, (L 993	Aenkes disease ± 309400
3COR	3CL6 Corepressor	Jg <i>Nat Genet</i> 36:411,ζL 2004	Aicrophthalmia, ±300166 yndromic 2

DLG3	Disc large, drosphilia, homologue	oPhilips <i>Orphanet J Rare</i> (L <i>)is</i> 9:49, 2014	Mental± 300850etardation,ζ-linked 90
FAM58A	<sup>7</sup> amily With Sequence Similarity 5 <i>M</i> ember A	583reen <i>J Med Genet</i> XL 3:594, 1996; Unger Nat Genet 40:287, 2008	STAR syndrome <sup>1</sup> 300707
7LNA	ilamin A	Kobertson     AJMG     AKL       40:1726, 2006     AKL	Aultiple OMIM 300017 lassifications
<i>ЪРСЗ</i>	Hypican 3	Cottereau AJMG C Semin(L Aed Genet 163:92, 2013	Simpson-Golabi-B <sup>1</sup> 312870 hmel syndrome, ype 1
41D1	Aidline 1	PreiksaitieneClin(LDysmorphol 24:7, 2015	)pitz GBBB! 300000 yndrome, type I
VAA10	J-Alpha -Acetyltransferase 1 JatA Catalytic Subunit	0,.enz Z KinderheilkdKL '7:384, 1955	Aicrophthalmia, <sup>1</sup> 309800 yndromic 1
VSDHL	JAD(P) Dependent Stero Dehydrogenase-Like	id&önig <i>J Am Acad</i> &L <i>Dermatol</i> 46:594, 2002	CHILD syndrome <sup>1</sup> 308050
ЧGA	Phosphatidylinositol Glycan Anch Biosynthesis Class A	orohnston <i>AJHG</i> 90:295,(L :012	Aultiple± 300868ongenitalnomalies-hypotoiia-seizuresyndrome 2
<i>PORCN</i>	'orcupine O-Acyltransferase	Suskan <i>Pediatr DermatoKL</i> ':283, 1990	<sup>7</sup> ocal dermal <sup>1</sup> 305600 1ypoplasia
MC1A	Structural Maintenance C Chromosomes 1A	DfDeardorff <i>GeneReviews</i> ® (L <i>Book Section Seattle(WA),</i> 993	Cornelia de Lange <sup>!</sup> 300590 yndrome 2
JPF3B	JPF3B, Regulator Of Nonsen Aediated MRNA Decay	se.ynch Eur J Med Genet(L 5:476, 2012	JA 300298
'IC3	Lic Family Member 3	Chung <i>AJMG</i> 155:1123, (L 2011	/ACTERL ± 314390 ssociation
)SR1	Odd-Skipped Related Transciptio	on'hang <i>Hum Mol Genet</i> Jnknown 20:4167, 2011	JA <sup>™</sup> 608891
PALB2	'artner and Localizer of BRCA2	Gia Nat Genet 39:159,Jnknown 2007	Fanconi Anemia, <sup>1</sup> 610832 Complementation Group N
PIK3CA	Phosphatidylinositol3-kinasCatalytic, Alpha	se\hmad Clin Dysmorphol 8:1, 2009	CLOVE syndrome <sup>1</sup> 612918
H2B1	H2B Adaptor Protein 1	Sampson <i>AJMG</i> Jnknown 52:2618, 2010	JA <sup>±</sup> 608937

ITNI	Atrophin 1	Palmer Am. J. Hum.AD Fenet. 104: 542-552, 2019	Congenital 07462 ypotonia, pilepsy, levelopmental lelay, and digital nomalies
33GLCT	3eta-3-glucosyltransferase	Dassie-Ajdid <i>Clin Genet</i> AR 90-492, 2009	Peter-Plus 10308 Syndrome
FANCD2	<sup>2</sup> anconi Anemia Group D2	Calb Am. J. Hum. GenetAR 30: 895-910, 2007	Vanconi anemia, 13984 omplementation group D2
<b>ANCE</b>	A Complementation Group E	Vegner <i>Clin. Genet.</i> 50:\R 79-482, 1996	Vanconi anemia, 13976 omplementation group E
CYP11B1	Cytochrome P450, Subfamily XI Polypeptide 1	B,Jlenthoj <i>Acta Endocr</i> . 93:\R/AD \4-99, 1980 ;	Adrenal iyperplasia, ongenital, due to 1-beta-hydroxyla e deficiency
DLL4	Delta Like Canonical Notch Ligar	ndöhutter <i>Genes Dev.</i> 14:\D 313-1318, 2000	Adams-Oliver i05185 yndrome 6
ИУМ2	Zinc Finger	Connaughton <i>Am. J. Hum</i> .\D <i>Fenet.</i> 107: 727-742, 2020	Jeurodevelopmen 02221 al-craniofacial yndrome with ariable renal and ardiac bnormalities
PTK7	Protein-tyrosine kinase	Chan <i>eLife</i> , 11, e74777,\R :022	osterior urethrab01890 alve
1 <i>FF3</i>	ALF Transcription Elongatio	onshimizu J. Hum. Genet.AD 4: 1041-1044, 2019	CINSSHIP 01464 yndrome
CFAP418	Cilia-and-Flagella-Associated Protein 418	teon <i>Hum. Molec. Genet</i> .AR 5: 2283-2294, 2016	Bardet-Biedl14477yndrome 21
CENPF	Centromeiric protein	Filges Hum. Mutat. 37:AR 559-363, 2016	stromme 500236 yndrome
CHRNA3	Cholinergic receptor, neuron icotinic, alpha polypeptide 3	al/ann <i>Am. J. Hum. Genet</i> .\R 05: 1286-1293, 2019	Bladder 18503 lysfunction, utonomic, with mpaired pupillary eflex and econdary CAKUT

<i>EBP</i>	3mopamil-binding protein	Ailunsky Am. J. Med.(LD/XLR Jenet. 116A: 249-254, 2003	Chondrodysplasia 00205 ounctata, X-linked lominant; MEND yndrome
ERCC4	RCC Excision repair indonuclease catalytic subunit	4, Jiedernhofer <i>Nature</i> 444: AR 038-1043, 2006	KFEprogeroid 33520yndrome
FANCG	<sup>7</sup> A Complementation Group G	le winter Nature Genet.AR 20: 281-283, 1998	anconi anemia, 02956 omplementation group G
HSPA9	leat-shock 70-KD protein 9	Royer-Bertrand <i>Sci. Rep</i> .\D/AR : 17154, 2015	Even-plus i00548 yndrome
CANSL1	CAT8 Regulatory NSL comple ubunit 1	ex,Cooleen <i>Nature Genet</i> .AD 8: 999-1001, 2006	Koolen-De Vries/12452 yndrome
COX14	Cytochrome c oxidase assemt actor COX14	olyVeraarpachai Am. J.AR Ium. Genet. 90: 142-151, :012	Aitochondrial 14478 omplex IV leficiency, nuclear ype 10
EDNRA	Endothelial receptor type A	Gordon Am. J. Hum.AD Genet. 96: 519-531, 2015	Aandibulofacial 31243 lysostosis with lopecia
FANCC	<sup>2</sup> A Complementation Group C	JA AR	anconi anemia, 13899 omplementation group C

*AR*, autosomal recessive; *AD*, autosomal dominant; *NA*, not available; *OMIM*, *Online Mendelian Inheritance in Man*; *XL*; *X-linked*; *#*, phenotype MIM number; Unknown, mode of inheritance not clearly characterized; \* gene/locus MIM number if not phenotype MIM number available.

Supplementary Table S5: genes that represent monogenic causes of human autosomal dominant tubulo-interstitial kidney disease, if mutated. (Sorted alphabetically by mode of inheritance).

Jene	Protein	Reference	Aode nheritano	of henotype ce	)MIM iumber
INF1B	Iepatocyte nuclear factor 2	Lindner <i>Hum Mol G</i> :4:263, 1999	Genet\D	Renal cysts liabetes syndrome	and <sup>!</sup> 137920 e
<i>AUC-1</i>	Aucin-1	Cirby Nat Genet 45: 2013	299,AD	Aedullary tidney disease 1	cystic <sup>!</sup> 174000
<i>₹EN</i>	Renin	'ivna <i>AJHG</i> 85: 204, 20	09 AD	Familial ju syperuricemic sephropathy 2	venile <sup>t</sup> 613092
EC61A1	EC61 complex, alpha 1	3olar <i>AJHG</i> 99:174, 201	6 AD	Pamilial ju Nyperuricemic Nephropathy 4	venile <sup>!</sup> 617056

JMOD	Jromodulin	łart	J	Med	<i>Genet</i> \D	amilial	juvenile <sup>1</sup> 162000
		9(12):8	882, 2	002		yperuricemic	
						ephropathy 1	

AD, autosomal dominant; OMIM, Online Mendelian Inheritance in Man; XL; X-linked; #, phenotype MIM number.

Supplementary Table S6: genes that represent monogenic causes of human chronic glomerulopathies, if mutated.

(Sorted alphabetically by mode of inheritance)

Jene	rotein	Reference	Aode on heritance	Phenotype	)MIM umber
IDAMTS13	ADAM netallopeptidase w hrombospondin type notif 13	Levy Nature 413: 488, 2001 ith e 1	AR	amilial thrombot hrombocytopenic ourpura	ic <sup>!</sup> 274150
COL4A4	Collagen type IV alp chain	bhaAochizuki Nat Genet 8:77, 1994	٨R	Alport syndrome	<sup>±</sup> 203780
EIF2AK3	Eukaryotic Translati nitiation Factor Alpha Kinase 3	ionDelepine Nature Genet 25:406 2:000	5,4R	Volcott-Rallison yndrome	<sup>£</sup> 226980
?LG	lasminogen	Schuster Blood 93:3457, 1999	AR	Plasminogen Deficiency, Type 1	<sup>!</sup> 217090
CFHR5	Complement factor elated 5	H3ale Lancet 376:794, 2010	٨D	Vephropathy due CFHR5 deficiency	to <sup>!</sup> 614809
7N1	ibronectin	Castelletti PNAS 105:2538, 2008	٨D	Bomerulopathy wi ibronectin deposits 2	th <sup>!</sup> 601894 2
<sup>7</sup> OXC2	Forkhead Box C2	(ildirim-TorunerAJMO31A:281, 2004	GAD	<i>symphedema-distich</i> <i>syndrome wi</i> <i>enal disease ar</i> <i>liabetes mellitus</i>	ia <sup>!</sup> 153400 th id
<del>3</del> SN	Jelsolin	Aaury FEBS Lett 260:85, 1990	٨D	Amyloidosis, Finnish ype	<sup>1</sup> 105120
.YZ	Jysozyme	Pepys Nature 362:553, 1993	٨D	Renal amyloidosis	<sup>!</sup> 105200
THBD	Thrombomodulin	Delvaeye NEJM 361:345, 2009	٨D	busceptibility typical hemolyt remic syndrome 6	to <sup>!</sup> 612926 ic
JLC37A4	Solute Carrier Fam 7 Glocose-6-Phosphat Pransporter), Mem	ilyVilson <i>JIMD Rep</i> 58:122, 2021 re ber	٨D	Congenital disorder ;lycosylation, typ Iw	of 619525

OX18	RY-Box 18	Sherwood Arch Dis ChildAD 52:1278, 1987	Iydrotrichosis-Lymph <sup>1</sup> 137940 dema-Telangiectasia- Renal Defect Syndrome
PRY2	Sprouty RT Signaling Antagonist 2	K <i>A</i> ilillo <i>Europ J Hum Genet</i> AD 2 :3:1673, 2015	Susceptibility to IgA <sup>1</sup> 616818 hephropathy 3
73	Complement C3	Fremeaux-Bacchi <i>Blood</i> 112:4948, AR/AD 2008	C3deficiency, 613779Susceptibilityto 612925iemolyticuremicyndrome, atypical 5
CD46	D46 molecule	Joris <i>Lancet</i> 362:1542, 2003 AR/AD	Atypical <sup>1</sup> 612922 nemolytic-uremic yndrome, type 2
CFB	CFB	Joicoechea de Jorge <i>PNAS</i> \R/AD 04:240, 2007	Complement factor B <sup>1</sup> 615561 leficiency, Susceptibility to nemolytic uremic <sub>1612924</sub> yndrome, atypical 4
CFH	Complement factor H	Edelsten Arch Dis Child 53:255, AR/AD 978	Complement factor H <sup>1</sup> 609814 leficiency, ±235400 Susceptibility to lemolytic uremic yndrome, atypical 1
CFHR1	Complement factor 1 elated 1	H <sup>2</sup> ipfel <i>PLoS</i> Genet 3:e41, 2007 AR/AD	Susceptibility to <sup>1</sup> 235400 nemolytic uremic yndrome, atypical
CFI	Complement factor I	Fremeaux-Bacchi <i>J Med Genet</i> 41:AR/ AD 84, 2004	Complement factor I <sup>1</sup> 610984 leficiency, Iemolytic uremic <sup>1</sup> 612923 yndrome, atypical
CFHR3	CFHR3	Lipfel PLoS Genet 3: e41, 2007 AR/ AD	Susceptibility to <sup>1</sup> 235400 emolytic uremic yndrome, atypical
COL4A3	Collagen type IV alph chain	na.emmink <i>Hum Mol Genet</i> 3:1269, \R/AD 994	Alport syndrome <sup>1</sup> 203780 Benign familial <sup>1</sup> 104200 Jematuria
COL4A5	Collagen type IV alph chain	naAntignac <i>J Clin Invest</i> 93:1195, KL 994	ζ-linked Alport <sup>1</sup> 301050 yndrome
COL4A6	Collagen type IV alph	natenieri Hum Mutat 4:195, 1994 (L	VA 303631

ONASE1L3	Deoxyribonuclease	Al-Mayouf Nature Genet. 43:AR	systemic lupus/02244
	-like-3	186-1188, 2011	rythematosus 16

Supplemental Table S7: Genes that represent monogenic causes of chronic kidney disease (other), if mutated.

Gene	Protein	Reference	Mode Inheritance	ofPhenotype	OMIM#
KYNU	Kynureninase	Christensen Inherit Metab 30:248, 2007	JAR Dis	Hydroxyknurenin	nuria 236800
LDHA	Lactate dehydrogenase A	Maekawa Am Hum Ge 39:232, 1986	JAR net	Glycogen sto disease XI	orage612933
MEFV	MEFV Inna. Immunity Regulator, Pyrin	teBenson Ann Int Med 87:31, 197	ernAR 7	Familial Mediterranean AR	249100 fever,
MMACHC	Metabolism Cobalamin Associated C	ofLerner-Ellis I Genet 38:93, 20	NatAR 106	Methylmalonic aciduria homocystinuria, type	277400 and cdlC
MMUT	Methylmalonyl-C A Mutase	o Ledley Bioess 12:335, 1990	ays AR	Methylmalonic aciduria, mut(0)	251000 type
MVK	Melcalonate Kinase	Prietsch Pediatrics 111:258, 2003	AR	Mevalonic acidu	ria 610377
PCBD1	Pterin-4-alpha-cc binolamine Dehydratase 1	nrFerrè J Am S Nephrol 25:5 2014	Soc AR 74,	Hyperphynylalar mia, BH4-Defia D	nine 264070 cient,
<i>PET100</i>	PET100 Cytochrome Oxidase Chaperone	Lim Am J H cGenet 94:2 2014	umAR 09,	Mitochondrial Complex Deficincy, Nu Type 12	619055 IV clear
SARS2	Seryl-tRNA Synthetase 2	Belostotsky An Hum Ge 88:193, 2011	n JAR net	Hyperuricemia, Pulmonary Hypertension, 1 Failure, Alkalosis Syndro	613845 Renal and me

SLC5A1	Solute Carrier Abdullah JAR Family 5Pediatr (Sodium/Glucose Gastroenterol Nut Cotransporter), 23:561, 1996 Member 1	Glucose/Galactose 606824 Malabsorption
UQCC2	Ubiquiinol-Cytoch Tucker PLoSAR rome C ReductaseGenet Complex Assembly9:e1004034, 2013 Factor 2	Mitochonrial 615824 Complex III Deficiency, Nuclear Type 7
NLRP3	NLR Family PyrinHoffman Am JAD Domain-ContaininHum Genet g 3 66:1693, 2000	Famalial cold120100 inflammatory syndrome 1
NLRP3	NLR Family PyrinDodé Am J HumAD Domain-Containin Genet 70:1498, g 3 2022	Muckle Wells191900 syndrome
NSD1	Nuclear Sotos N Engl JAD Receptor-binding Med 271:109, set domain protein1964 1	Sosto Syndrome 117550
SLC47A1	Solute CarrierGreenberg Am JAD Family 47,Med Genet Member 1 62:247, 1996	Smith-Magenis 182290 Syndrome
STX16	Syntaxin 16 Levine Am J MedAD 74:545, 1983	Pseudohypoparathyro603233 idism Type IB
PGK1	Phosphoclycerate Rosa Blood 60:84,XLR Kinase 1 1982	Phosphoglycerate 300653 Kinase 1 Deficiency
WAS	Wasp ActinStanden Q J MedXLR Nucleation 59:401, 1986 Promoting Factor	Wiskott-Aldrich 301000 Syndrome
PLA2R1	Phospholipase A2Debiec N Engl JUnknown Receptor 1 Med 346:2053	Membranous 614692 Nephropathy
SACS	Sacsin Criscuolo AR Neurology 62: 100-102, 2004	Spastic ataxia, 604490 Charlevoix-Saguenay type
UQCRQ	Ubiquinol-cytochr Barel Am. J. Hum.AR ome c reductase, Genet. 82: complex III 1211-1216, 2008 subunit VII	Mitochondrial 612080 complex III deficiency, nuclear type 4
ADA2	Adenosine Zhou New Eng. J.AR deaminase Med. 370: 911-920, 2014	Vasculitis, 607575 autoinflammation, immunodeficiency, and hematologic defects syndrome

<i>C5</i>	Complement component 5	Nishimura NewAD/AR Eng. J. Med. 370: 632-639, 201	Eculizumab, poor120900 response to
FOXP3	Forkhead box P3	Misra GeneXLR 25;581(1):57-65, 2016	Susceptibility to 300292 ESRD
HPS1	HPS1 Biogeness of lysosome organelles complex3, subun l	isO'Brian JournalAR 11of internal medicine, 290(1), it129–140, 2021	Hermansky-Pudlak 604982 syndrome 1; renal failure
ITGA6	Integrin, alpha 6	Ruzzi J. Clin.AR Invest. 99: 2826-2831, 1997	Epidermolysis 147556 bullosa, junctional 6, with pyloric atresia
KCNA1	Potassium channel, voltage-gated, shaker-related subfamily, membe l	Chan AD Neurogenetics 8: 131-135, 2007 er	Episodic 176260 ataxia/myokymia syndrome
SDHB	Succinate dehydrogenase complex, subun B, iron sulfi protein	Fairchild JAMAAD 242: 2210-2211, it1979 ır	Pheochromocytoma 185470
SDHD	Succinate dehydrogenase complex, subun D, integra membrane proteir	Fairchild JAMAAD 242: 2210-2211, it1979 al 1	Pheochromocytoma 602690
TMEM127	Transmembrane protein 27	Fairchild JAMAAD 242: 2210-2211, 1979	Pheochromocytoma 613403
CACNA1H	Calcium channe voltage-dependen T type, alpha-1 subunit	l,Scholl eLife 4:AD t,e06315, 2015 H	Hyperaldosteronism, 607904 familial, type IV
CCND1	Cyclin D1	Yu The Journal of AD urology, 172(6 Pt 1), 2410–2413, 2004	Von Hippel-Lindau168461 Syndrome
CLCN2	Chloride channe 2	elScholl NatureAD Genet. 50: 349-354, 2018	Hyperaldosteronism, 600570 familial, type II

CYP11B2	Cytochrome P450, Chen ToxicologyAR subfamily XIB, and applied polypeptide pharmacology, 279(2), 95–102, 2014	Hypoaldosteronism, 124080 congenital, due to CMO I deficiency / Hypoaldosteronism, congenital, due to CMO II deficiency
HNF1A	HNF1 HomeoboxReibouissou Hum.AD/AR A Molec. Genet. 14: 603-614, 2005	Renal cell carcinoma 142410
KCNJ18	Potassium Ryan Cell 140:AD channel, inwardly88-98, 2010 rectifying, subfamily J, member 18	Thyrotoxic periodic613236 paralysis, susceptibility to, 2
FGF3	Fibroblast growthVasei CancerAR factor 3 Genet Cytogenet 194:88-95, 2009	Wilms tumour 164950
UBE3A	Ubiquitin-protein Wu Genet Mol ResAD ligase E3A 15:gmr15049023, 2016	Angelman syndrome 601623
NTN1	Netrin-1 Ranganathan AD Mediators Inflamm 2014:525891, 2014	Acute kidney601614 injury/CKD
PERM1	PPARGC1- andN/A AD ESRR-induced regulator muscle, 1	N/A 615921
COA8	Cytochrome CN/A AD Oxidase Assembly Factor	N/A N/A
COX20	Cytochrome cSzklarczyk Hum.AR oxidase assemblyMolec. Genet. 22: factor COX20 656-667, 2013	Mitochondrial 614698 complex IV deficiency, nuclear type 11; lactic acidosis
COX8A	Cytochrome cHallman BrainAR oxidase, subunit139: 338-345, 8A 2016	Mitochondrial 123870 complex IV deficiency, nuclear type 15
FANCM	FAKim Genes (Besel)ARComplementation17;12(5):7511,Group M2021	Hypertensive kidney609644 disease

FASTKD2	Fast kinase Ghezzi Am. J.AR domain 2 Hum. Genet. 83: 415-423, 2008	Combined oxidative612322 phosphorylation deficiency 44
KCNK3	Potassium Ma New Eng. J.AD channel, subfamilyMed. 369: K, member 3 351-361, 2013	Pulmonary 603220 hypertension, primary, 4
BMPR2	Bone Rigelsky Am. J.AD morphogenetic Med. Genet. protein receptor,146A: 2551-2556, type II 2008	Pulmonary 600799 hypertension, familial primary, 1, with or without HHT; Pulmonary hypertension, primary, fenfluramine or dexfenfluramine-asso ciated; Pulmonary venoocclusive disease 1
HBB	Hemoglobin—beta Monk PrenatalAD locus Diag. 13: 45-53, 1993	Sickle cell anemia 141900
HSD3B2	3-Beta-hydroxyste Bongiovanni Clin.AR roid Invest. 41: dehydrogenase 2 2086-2092, 1962	Adrenal hyperplasia, 613890 congenital, due to 3-beta-hydroxysteroid dehydrogenase 2 deficiency

Supplementary Table S8: Genes that represent monogenic causes of human diabetic kidney disease, if mutated.

Gene	Protein	Reference	Mode Inheritance	ofPhenotype	OMIM#
<i>C</i> 7	Compliment component 7	Chen J Biochem 12. 2021	CellAR 3:481,	DKD	
CCR2	Chemokine, CC Mot Receptor 2	if,Chen J Biochem 12. 2021	CellAR 3:481,	DKD	

GPR158	<i>G Protein-coupled recepto</i> 158	rSandholm Diabetologia 65:1495, 2022	AR	DKD
LSM14A	LSM14A mRNA Processing Body Assembly Factor	gSandholm Diabetologia 65:1495, 2022	AR	DKD
MFF	Mitochondrial Fission Factor	nSandholm Diabetologia 65:1495, 2022	AR	DKD
MOXD1	Monooxygenase, DBH-lik 1	eSandholm Diabetologia 65:1495, 2022	AR	DKD
PYCARD	PYD and Care Domain-containing Protein	dChen J 1 Biochem 123 2021	CellAR :481,	DKD
COL1A2	Collagen of Skin, Tendon and Bone, Alpha-2 Chain	n,Chen J Biochem 123 2021	CellAD :481,	DKD
COL6A3	Collagen Type 6, Alpha-3	Chen J Biochem 123 2021	CellAD :481,	DKD
COL20A1	Collagen Type XX, Alpha-i	Sandholm Diabetologia 65:1495, 2022	AD	DKD
DCLK1	Doublecortin-like Kinase I	Sandholm Diabetologia 65:1495, 2022	AD	DKD
EIF4E	Eukaryotic Translation Initiation Factor 4E	nSandholm Diabetologia 65:1495, 2022	AD	DKD
PAX4	Paired Box Gene 4	Plengvidhya J Endocrinol M 92:2821, 2007	Clin Ietab	MODY type IX 612225

NEUROD1	Neurogenic Differentiat 1	ionMalecki Nat GenetUnknown 23:323, 1999	MODY 6 60	6394
PDX1	Pancreas/Duodenum Homeobox Protein 1	Fajans N Engl JUnknown Med 345:971, 2001	MODY type IV 60	6392
GREM1	Gremlin 1 homolog, cyst knot superfamily	tineMcKnight JournalAR of the American Society of Nephrology : JASN, 21(5), 773–781, 2010	Diabetic 60. nephropathy	3054
CEL	Carboxyl-ester lipas	Raeder Nature AD Genet. 38: 54-62, 2006	Maturity-onset 11- diabetes of the young, type VIII	4840
CYP11A1	Cytochrome P4 subfamily XIA, polypept 1	50,Pagotto Mol CellAR tideEndocrinol 15:111170, 2021	Adrenal 116 insufficiency, congenital, with 46XY sex reversal, partial or complete	8485
GCK	Glucokinase	Njolstad New Eng.AD/AR J. Med. 344: 1588-1592, 2001	Diabetes 13 mellitus, noninsulin-depe ndent, late onset; Diabetes mellitus, permanent neonatal 1	8079

INS	Insulin	N/A	AD/AR	Diabetes 176730 mellitus
				insulin-depende
				nt. 2: Diabetes
				mellitus,
				permanent
				neonatal 4;
				Hyperproinsulin
				emia;
				Maturity-onset
				diabetes of the
				young, type 10

KCNJ11	Potassium channel, Yorifuji J. Clin inwardly rectifyingEndocr. Metab. 90:	AD/AR Diabetes 600937 mellitus,
	subfamily J, member 11 3174-3178, 2005	transient
		neonatal 3;
		Diabetes,
		permanent
		neonatal 2, with
		or without
		neurologic
		features;
		Hyperinsulinemi
		c hypoglycemia,
		familial, 2;
		Maturity-onset
		diabetes of the
		young, type 13;
		Diabetes
		mellitus, type 2,
		susceptibility to

KLF11	Kruppel-like-factor 11	Neve	Proc.	Nat.Maturity-onset Maturity-onset 603301
		Acad.	Sci.	102: diabetes of the diabetes of the
		4807-4	812, 200	5 young, type VII young, type VII

Supplementary Table S9: genes that represent monogenic causes of hereditary amyloidosis, if mutated. (Sorted alphabetically by mode of inheritance).

Jene	Protein	<i>leference</i>	Aode nheritance	oPhenotype	)MIM iumber
APOC2	Apolipoprotein C2	Cufova J Pathol 71:687, 2	<i>Clin</i> \R 2018	Apolipoprotein Deficiency	C-II:07750
APOA1	Apolipoprotein A1	Cufova J Pathol 71:687, 2	<i>Clin</i> \D 2018	Amyloidosis, 3 ypes	or more 05200
APOA2	Apolipoprotein A2	Cufova J Pathol 71:687, 2	<i>Clin</i> 2018	Iereditary amylo	vidosis
APP	Amyloid beta Precurso Protein	orKufova J Pathol 71:687, 2	<i>Clin</i> \D 2018	Cerebral ngiopathy, PRN	amyloid/05714 P-related
32M	3eta-2-Microglobulin	Cufova J Pathol 71:687, 2	<i>Clin</i> \D 2018	Amyloidosis, risceral	familial 05200
CST3	Żystatin C	Cufova J Pathol 71:687, 2	<i>Clin</i> \D 2018	Cerebral Angiopathy, CST	Amyloid 05150 '3-related
GA	ibrinogen Alpha Chai	n Cufova J Pathol 71:687, 2	<i>Clin</i> AD 2018	Amyloidosis, risceral	familial 05200

3SN	Jelsolin	Cufova <i>J Clin</i> AD Pathol 71:687, 2018	Amyloidosis, Finnish 05120 Type
TM2B	ntegral Membran Protein 2B	nečufova J Clin Pathol 71:687, 2018	Amyloidosis
XZ	.ysozyme	Cufova <i>J Clin</i> AD Pathol 71:687, 2018	Amyloidosis, renal 05200
'RNP	Prion Protein	Cufova <i>J Clin</i> AD Pathol 71:687, 2018	Cerebral amyloid 37440 ngiopathy, PRNP-related
TR	Transthyretin	Cufova <i>J Clin</i> AD Pathol 71:687, 2018	Amyloidosis, hereditary, 05210 ransthyretin-related
32M	3eta-2-microglobulin	/alleix New Eng. JAD     Aed.   366:     276-2283, 2012	Amyloidosis, familial 09700 risceral
GA	ibrinogen, A Alpl olypeptide	ha3enson <i>Nature</i> AD <i>Fenet.</i> 3: 252-255, 993	Amyloidosis, familial 34820 risceral